

Targeting the hallmarks of aging: preclinical evidence and molecular mechanisms underlying the therapeutic potential of *Centella asiatica* (L.) Urban

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Abstract

As the global demographic shifts toward an aging population, the escalating prevalence of age-related pathologies necessitates the identification of geroscience-based interventions. *Centella asiatica* (L.) Urban has emerged as a potent candidate for healthspan extension due to its dense concentration of bioactive pentacyclic triterpenoids, specifically asiaticoside, madecassoside, asiatic acid, and madecassic acid. This review synthesizes preclinical evidence demonstrating the capacity of *C. asiatica* to pharmacologically modulate the fundamental hallmarks of aging. At the molecular level, its constituents have been shown to maintain genomic stability by enhancing DNA repair mechanisms and mitigating epigenetic drift through the regulation of histone modifications. Furthermore, the plant exerts profound effects on mitochondrial bioenergetics, restoring mitochondrial homeostasis to counteract age-related metabolic decline. Analysis of *in vitro* and *in vivo* models reveals that *C. asiatica* suppresses the senescence-associated phenotype and systemic "inflammaging". We further detail the plant's neuroprotective and tissue-regenerative properties, driven by the neutralization of reactive oxygen species (ROS) and the activation of antioxidant signaling. By systematically targeting these discrete cellular aging drivers, *C. asiatica* represents a potent, naturally-derived scaffold for the development of multi-target therapies aimed at extending biological healthspan. This review underscores the necessity of translating these preclinical mechanistic insights into targeted gerotherapeutic strategies.

Keywords: *Centella asiatica*, aging, neuroprotection, anti-inflammation, triterpenoids

1. Introduction

The aging of the global population has become a critical challenge in modern medicine. By 2050, individuals aged over 60 years are projected to reach two billion, representing 21% of the global population (United Nations, 2013). However, increased life expectancy is often marred by a period of late-life morbidity. Consequently, there is a profound gap between

lifespan and healthspan, the period of life spent in good health, free from the chronic infirmities of age-related diseases (ARDs) (Shlisky *et al.*, 2017).

The burgeoning field of geroscience posits that aging itself is the primary risk factor for most chronic pathologies, including neurodegenerative, cardiovascular, and metabolic disorders (Niccoli and Partridge, 2012). Rather than treating these conditions in isolation, targeting the underlying biological drivers of aging offers a more holistic therapeutic approach. These drivers are categorized within the "Hallmarks of Aging" framework, which describes a complex interplay of molecular and cellular malfunctions (López-Otín *et al.*, 2023). These hallmarks are classified into (i) primary hallmarks, such as genomic instability, telomere attrition, epigenetic alterations, and loss of proteostasis, which represent the initial triggers of cellular damage; (ii) antagonistic hallmarks, including cellular senescence and mitochondrial dysfunction, which arise as compensatory or defensive responses to damage but ultimately become deleterious; and (iii) integrative hallmarks, like stem cell exhaustion and "inflammaging", a state of chronic, low-grade systemic inflammation which collectively result in the functional decline observed in aging phenotypes (López-Otín *et al.*, 2023). Understanding these interconnected mechanisms will shed light on strategies to extend healthspan (Schmauck-Medina *et al.*, 2022).

In the search for natural bioactive compounds capable of modulating these hallmarks, *Centella asiatica* (L.) Urban (CA), a medicinal herb with a long history in Ayurvedic and Traditional Medicine (Sun *et al.*, 2020), has gained significant attention. The therapeutic efficacy of CA is largely attributed to its unique profile of pentacyclic triterpenoids, most notably asiaticoside, madecassoside, asiatic acid, and madecassic acid (James and Dubery, 2009). While CA has been studied for its wound-healing and cognitive-enhancing properties, recent pre-clinical investigations have begun to unravel its deeper role as a multi-target gerotherapeutic. These studies suggest that CA does not merely mask symptoms but interferes with the aging process at the subcellular levels.

Despite the promising data, there is a critical need to distinguish between the efficacy of whole botanical extracts and isolated compounds, as well as the varying degrees of evidence provided by different experimental models. This review provides a comprehensive synthesis of the evidence supporting the use of *C. asiatica* in healthspan extension. We aim to interpret the molecular pathways through which CA modulates the hallmarks of aging and to identify the gaps remaining for future clinical translation.

Table 1. Summary of the multi-target mechanisms of triterpenoids of *C. asiatica* in regulating the hallmarks of aging. Evidence levels are categorized as "direct" (studies specifically focusing on aging models) or "indirect" (mechanistic studies in non-aging contexts).

Hallmarks - evidence level	Mechanisms	Models
Total extracts		
Proteostasis - indirect	Reducing amyloid-beta levels in Alzheimer's-related brain pathology (Dhanasekaran <i>et al.</i> , 2009)	<i>In vitro</i>
Others - direct	Enhancing learning performance and cognitive flexibility in aged mice (Hack <i>et al.</i> , 2025)	<i>In vivo</i>
Asiaticoside		
Genomic instability - indirect	Reducing radiation-induced DNA strand breaks (Shen <i>et al.</i> , 2020)	<i>In vitro</i> , <i>In vivo</i>
mitochondrial bioenergetics - indirect	Enhancing mitochondrial respiration and ATP production (Boondam <i>et al.</i> , 2025)	<i>In vitro</i> , <i>in vivo</i>
Cellular senescence - direct	Delaying senescence and decreases ROS generation by modulating the TGF-β1/Smad pathway (Jiang <i>et al.</i> , 2022).	<i>In vitro</i>

'Inflammaging' - indirect	Reducing IL-6, TNF- α and IL-1 β via HO-1 upregulation, cAMP/PKA modulation and NF- κ B inhibition, thereby attenuating inflammation and apoptosis across arthritis, diabetic and LPS-induced injury models (He <i>et al.</i> , 2024; Luo <i>et al.</i> , 2022; Zhang <i>et al.</i> , 2022).	<i>In vivo</i> , <i>In vitro</i>
Others - indirect	Predicted to regulate synaptic plasticity via TSC1 - Rheb - FMRP and the AKT/mTORC1 pathway (Ibrahim <i>et al.</i> , 2023).	<i>In silico</i>
Madecassoside		
Proteostasis - indirect	Restoring synaptic and circadian function in PIMT deficits (Ling <i>et al.</i> , 2024).	<i>In vivo</i>
Autophagy - indirect, direct	Activates AMPK/autophagy to reduce ER stress and fatty liver (Choi <i>et al.</i> , 2023).	<i>In vivo</i> , <i>In vitro</i>
	Enhancing autophagic flux and lysosomal function to limit protein aggregation and oxidative stress (Ling <i>et al.</i> , 2017).	<i>In vitro</i>
Mitochondrial bioenergetics - indirect	Inducing ROS-mediated apoptosis and G2/M arrest, suppressing proliferation and migration via MAPK and PI3K/AKT (Hou <i>et al.</i> , 2025).	<i>In vitro</i>
Senescence - indirect	Suppressing NF- κ B signaling and decreases IL-6, TNF- α , and IL-1 β secretion, alleviating chronic inflammation associated with SASP and cellular aging (Won <i>et al.</i> , 2010; Yun <i>et al.</i> , 2008).	<i>In vitro</i>
'Inflammaging' - indirect	Inhibiting inflammation in <i>Propionibacterium acnes</i> and UV models, improving hydration and reducing melanogenesis (Shen <i>et al.</i> , 2019).	<i>In vitro</i>
Intercellular communication - indirect	Inhibiting acetylcholinesterase, reducing neuroinflammation and oxidative stress while stabilizing neuronal communication (Li <i>et al.</i> , 2025).	<i>In vitro</i>
Antioxidant - indirect	PRDX2-mediated redox control limits neurodegeneration (Zhong <i>et al.</i> , 2025).	<i>In vivo</i>
	Improving skin aging parameters by antioxidant mechanisms, enhancing collagen elasticity, hydration, and dermal integrity (Haftak <i>et al.</i> , 2008)	<i>Clinical</i>
Asiatic Acid		
Proteostasis - direct, indirect	Maintaining proteostasis and delaying cellular aging (Wang, 2014).	<i>In vitro</i>
	Improves cognition via proteostasis and hippocampal neurogenesis (Umka Welbat <i>et al.</i> , 2016). Modulating PI3K/Akt/mTOR, MAPK, and GSK3 β signaling to promote cell survival (Nataraj <i>et al.</i> , 2017; Wei <i>et al.</i> , 2018).	<i>In vivo</i>
Autophagy - indirect	Enhancing mitophagy, glycopagy; preserving mitochondrial homeostasis; reducing ROS (Qiu <i>et al.</i> , 2022; Yi <i>et al.</i> , 2020; Zhu <i>et al.</i> , 2021). Promoting autophagy to clear protein aggregates and protects neurons from proteotoxic stress (Yi <i>et al.</i> , 2020).	<i>In vitro</i> , <i>In vivo</i>
Mitochondrial bioenergetics - indirect, direct	Maintaining mitochondrial function and bioenergetics (Hu <i>et al.</i> , 2020). Inhibiting ROS-mediated apoptosis via Bcl-2/Bax modulation, reducing cytochrome c release and caspase activation (Hu <i>et al.</i> , 2020). Suppressing ROS-dependent apoptosis via Bcl-2/Bax modulation, reduced cytochrome c release, and caspase suppression in non-malignant cells (Tang <i>et al.</i> , 2009).	<i>In vitro</i>

	Repressing p38/JNK MAPK signaling and modulates AKT-mediated survival pathways (Yi <i>et al.</i> , 2022). Inducing $\Delta\Psi_m$ dissipation and ATP depletion via UCP/ANT involvement, triggering intrinsic apoptosis in cancer cells (Lu <i>et al.</i> , 2016).	
Senescence - indirect, direct	Delaying cellular senescence and reduces UV-induced ROS via modulation of the TGF- β 1/Smad signaling pathway, contributing to protection against photoaging (Dang <i>et al.</i> , 2024; Yun <i>et al.</i> , 2008).	<i>In vitro</i>
'Inflammaging' - indirect	Suppressing neuroinflammation in BV2 microglial cells via modulation of the Sirt1/NF- κ B signaling pathway (Qian <i>et al.</i> , 2018; Yuyun <i>et al.</i> , 2018).	<i>In vitro</i>
Intercellular communication - indirect	ERK activation drives osteogenic differentiation (Thammium <i>et al.</i> , 2023).	<i>In vitro</i>
Antioxidant - direct, indirect	Protects mitochondria against rotenone-induced apoptosis (Nataraj <i>et al.</i> , 2017). Protecting HepG2 cells from oxidative stress and apoptosis Via AKT/ERK (Papaiahgari <i>et al.</i> , 2006; Qi <i>et al.</i> , 2017). Scavenging ROS and improving oocyte quality and embryo development (Hu <i>et al.</i> , 2020; Qi <i>et al.</i> , 2021).	<i>In vitro</i>
	Restoring redox balance, and reducing oxidative stress - induced neuronal apoptosis in Alzheimer's models (Gray <i>et al.</i> , 2017; Rather <i>et al.</i> , 2018).	<i>In vitro</i> , <i>In vivo</i>
Madecassic acid		
'Inflammaging' - indirect	Reducing SASP-related inflammatory cytokines through NF- κ B inhibition (Gray <i>et al.</i> , 2017; Liu <i>et al.</i> , 2012; Rather <i>et al.</i> , 2018).	<i>In vitro</i> , <i>In vivo</i>
	Attenuating inflammatory mediator production and cartilage degradation in chondrocyte and osteoarthritis models (Fu <i>et al.</i> , 2022).	<i>In vitro</i> , <i>In vivo</i>
Intercellular communication - indirect	Activating PPAR γ /AMPK/ACC1 signaling, restoring Th17/Treg balance and suppressing pro-inflammatory cytokine production (Xu <i>et al.</i> , 2017).	<i>In vivo</i>
	Inflammasome inhibition promotes repair and remyelination (Li <i>et al.</i> , 2025).	<i>In vitro</i> , <i>In vivo</i>
Other components		
Intercellular communication - indirect	Araliadiol promotes hair growth via p38/PPAR- γ signaling (Park <i>et al.</i> , 2024).	<i>In vitro</i>
Others - direct	Caffeoylquinic acids ameliorate age-related cognitive changes in aged mice (Hack <i>et al.</i> , 2025).	<i>In vivo</i>
Antioxidant - indirect	Flavonoids reduce ROS levels, and protect skin cells from oxidative stress (Kim <i>et al.</i> , 2023).	<i>In vitro</i>
Antioxidant - indirect	Polyphenols reduce oxidative stress via radical scavenging and metal reduction (Ponnusamy <i>et al.</i> , 2008).	<i>In vivo</i>

2. Literature Review Approach / Review Methodology

Literature Search Strategy

A comprehensive literature search was conducted to identify relevant preclinical studies. We searched three primary electronic databases, including PubMed, ScienceDirect, and SpringerLink, covering until 03/2026.

The search utilized combinations of the following Boolean operators and keywords:

"*Centella asiatica*" OR "Gotu Kola" AND "hallmarks of aging"

"*Centella asiatica*" OR "Gotu Kola" AND "genomic instability"

"*Centella asiatica*" OR "Gotu Kola" AND "safeguarding proteostasis"

"*Centella asiatica*" OR "Gotu Kola" AND "enhancing autophagy"

"*Centella asiatica*" OR "Gotu Kola" AND "cellular senescence"

"*Centella asiatica*" OR "Gotu Kola" AND "inflammaging"

"*Centella asiatica*" OR "Gotu Kola" AND "intercellular communication"

"*Centella asiatica*" OR "Gotu Kola" AND "antioxidant"

Inclusion and Exclusion Criteria

Inclusion was limited to:

- (1) Peer-reviewed primary research articles.
- (2) Studies specifically investigating the molecular pathways of its triterpenoids related to the nine hallmarks of aging.
- (3) Articles published in the English language.

Exclusion criteria included clinical trial protocols without results, conference abstracts, and studies focusing solely on the botanical cultivation of the plant rather than its biological activity.

3. *Centella asiatica* and the Hallmarks of Aging

3.1. Primary hallmarks

The primary hallmarks of aging represent the foundational triggers of cellular damage, beginning with the loss of genomic stability. The genome is constantly exposed to exogenous and endogenous insults, including replication errors and oxidative stress, which cause mutations and chromosomal alterations. This leads to genomic mosaicism (López-Otín *et al.*, 2023; Vijg and Suh, 2013) and eventually contributes to aging and ARDs (Vijg & Dong, 2020). While endogenous DNA repair machinery such as base excision repair and nucleotide excision repair maintains genomic integrity, its functional efficacy diminishes significantly with age. This age-related attrition facilitates the accumulation of unrepaired lesions, driving genomic instability (Miller *et al.*, 2021). Closely linked to this genomic decay is the progressive attrition of telomeres. Telomere length declines with each division, triggering senescence or apoptosis at a critical threshold (Blackburn *et al.*, 2015; López-Otín *et al.*, 2023). As most somatic cells lack telomerase, telomeres undergo continuous attrition (Blasco, 2005; Chakravarti *et al.*, 2021). While this limits malignancy, excessive erosion impairs tissue regeneration and accelerates aging, marking it as a hallmark distinct from genomic instability (López-Otín *et al.*, 2023). Notably, telomerase reactivation can reverse aging phenotypes, identifying telomere dynamics as a key target for healthspan interventions (Jaskelióff *et al.*, 2011). Beyond the physical structure of the DNA sequence, epigenetic dysregulation further compounds cellular dysfunction. Shifts in DNA methylation patterns (the "epigenetic clock"), aberrant histone modifications, and the reactivation of transposable elements (retrotransposons) all contribute to the loss of cell identity and the progression of ARDs (Gorbulnova *et al.*, 2021; López-Otín *et al.*, 2023).

The convergence of genomic instability, telomere attrition, and epigenetic dysregulation creates a molecular foundation for biological aging. However, the inherent plasticity of these

primary hallmarks suggests that biological decline is not an unalterable trajectory, but rather a process susceptible to pharmacological attenuation. By enhancing the efficiency of DNA repair enzymes, stabilizing telomerase activity, and restoring histone acetylation/methylation patterns, geroscience-based interventions aim to preserve the structural and functional integrity of the cell. Preclinical evidence indicates that modulating these upstream drivers can effectively delay the transition from healthy aging to symptomatic disease. Consequently, targeting this triad of genetic and epigenetic integrity offers a potent strategy for extending healthspan, providing a molecular basis for the therapeutic application of multi-target botanicals like *C. asiatica* (Figure 1).

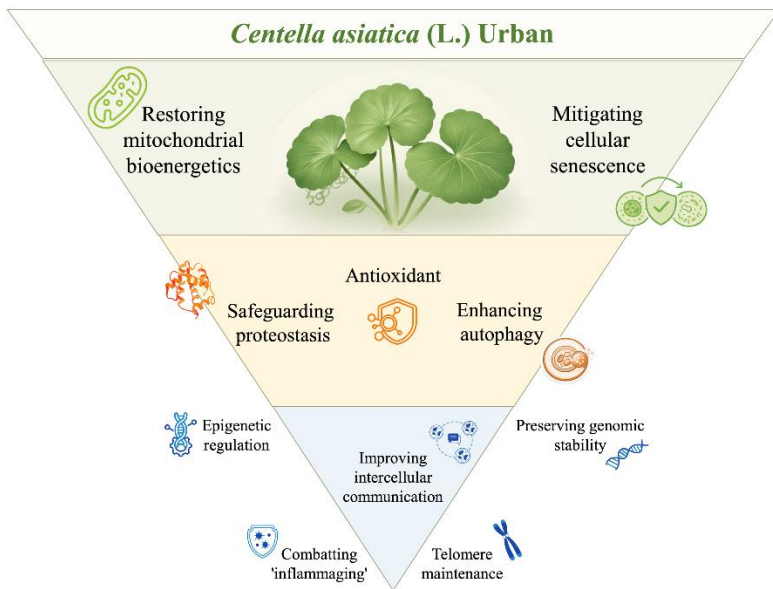


Figure 1. Multi-target effects of *Centella asiatica* on the hallmarks of aging. The hallmarks are organized hierarchically, with a top-to-bottom gradient representing the strength of evidence from direct aging-related studies. *C. asiatica* exerts prominent effects on mitochondrial function and cellular senescence, followed by antioxidant activity, proteostasis maintenance, and autophagy promotion. Direct evidence remains nascent for genomic stability, epigenetic regulation, intercellular communication, telomere maintenance, and aging-related chronic inflammation. Illustration created with Adobe Illustrator.

The anti-aging potential of *Centella asiatica* has been demonstrated across multiple experimental systems. A bioactive extract, DLBS1649, has been shown to preserve cellular replicative capacity by maintaining telomere length and modulating the expression of aging-associated genes, including telomerase reverse transcriptase (TERT), sirtuin 1 (SIRT1), and Klotho. In addition, the extract reduced lipid accumulation, suggesting a calorie restriction-mimetic effect. These cellular findings were further supported by in vivo studies in *Drosophila melanogaster*, where DLBS1649 treatment significantly extended lifespan, highlighting its potential to modulate physiological aging processes (Karsono *et al.*, 2021). These systemic

impacts are further elucidated through epigenetic profiling. For instance, water extracts of *C. asiatica* have been shown to modulate peripheral blood DNA methylation in aged mice, specifically targeting genes linked to antioxidant responses, metabolic regulation, and circadian rhythms (Monestime *et al.*, 2025).

Complementing these systemic effects, specific compounds like asiaticoside provide targeted protection against genotoxic stress by significantly reducing radiation-induced DNA strand breaks and improving cellular resistance to damage (Joy *et al.*, 2015). This protective efficacy extends to specialized tissues like the skin, where *C. asiatica* works synergistically within multi-component botanical formulations. When combined with extracts of *Punica granatum*, *Glycyrrhiza glabra*, and *Cynara scolymus*, it targets the extracellular matrix by inhibiting elastase and collagenase, the primary enzymes responsible for structural protein degradation, while simultaneously stimulating type I collagen synthesis in human dermal fibroblasts (Quiles *et al.*, 2022).

In summary, *Centella asiatica* emerges as a potent multi-targeted agent in the field of geroscience, addressing several fundamental hallmarks of aging (Figure 1, Table 1). By integrating mechanisms that preserve genomic stability, modulate the epigenetic landscape, and maintain the integrity of the extracellular matrix, it offers a comprehensive approach to both systemic longevity and localized tissue regeneration. Whether through the regulation of longevity-related genes like SIRT1 or the direct stimulation of structural proteins, *C. asiatica* demonstrates a unique dual-action capability—protecting existing biological structures from environmental and chronological decay while actively promoting the synthesis of youthful cellular components.

3.2. Proteostasis and autophagy

Proteostasis disruption is a hallmark of accelerated aging and neurodegeneration, characterized by the intra- and extracellular accumulation of misfolded or damaged proteins (Gerashchenko *et al.*, 2021; Hipp *et al.*, 2019). This collapse is largely driven by the age-associated impairment of the unfolded protein response (Maruthiyodan *et al.*, 2026), ubiquitin-proteasome system (UPS), and autophagic-lysosomal pathways, all of which exacerbates systemic proteotoxic stress (Hetz *et al.*, 2020).

Meanwhile, autophagy, a highly conserved degradation pathway via lysosome, responsible for degrading damaged organelles and proteotoxic aggregates, is defined by the decreased expression of core autophagy-related genes (ATGs), such as ATG5, ATG7, ATG12, beclin 1 (BECN1), unc-51 like autophagy activating kinase 1 (ULK1), and microtubule-associated protein 1A/1B-light chain 3 (MAP1LC3) (Lipinski *et al.*, 2010). This genetic decline is compounded by the depletion of endogenous autophagy-inducing metabolites, such as spermidine, and the dysregulation of regulatory signaling pathways like E1A-binding Protein p300 (EP300) (Alsaleh *et al.*, 2020; Xu and Wan, 2023; Zhang *et al.*, 2019).

Regarding *in vitro* mechanisms, *C. asiatica* and its bioactive compounds safeguard proteostasis by modulating key autophagic and proteolytic pathways. Asiatic acid serves as a potent inhibitor of advanced glycation endproducts (AGEs) (Wang, 2014), normalizes dysregulated autophagy in cardiac models by regulating LC3-II, BECN1, and p62 (Franceschi *et al.*, 2018). Furthermore, it induces apoptosis in various cancer models via PI3K/Akt/mTOR inhibition (Hao *et al.*, 2018; Ren *et al.*, 2016). Similarly, asiaticoside and madecassoside function as potent mTOR inhibitors (Zulkipli *et al.*, 2020), which enhance the autophagic clearance of toxic α -synuclein aggregates through dual AMPK activation and mTOR suppression, thereby restoring lysosomal function (Limanaqi *et al.*, 2019).

In *in vivo* models, these proteostatic effects translate into broad neuroprotective and systemic benefits. Asiatic acid restores redox balance and promotes neuronal survival in Parkinson's disease and myocardial injury models by modulating mitophagy- and glycopagy-based energy metabolism (Franceschi *et al.*, 2018; Nataraj *et al.*, 2017; Qiu *et al.*, 2022). Its systemic reach is evidenced by its ability to protect cognitive function in valproic acid-induced models

(Umka Welbat *et al.*, 2016), attenuates liver fibrosis through Bcl-2/Bax signaling (Wei *et al.*, 2018), and also improves diabetic nephropathy by restoring podocyte autophagy (Ni *et al.*, 2025). Madecassoside specifically alleviates deficits from protein-L-isoaspartate O-methyltransferase (PIMT) deficiency, restoring synaptic function disrupted by protein misrepair (Ling *et al.*, 2024). Furthermore, whole extracts, including water extracts and phytosomal preparations, selectively decrease hippocampal β -amyloid levels (Dhanasekaran *et al.*, 2009; Matthews *et al.*, 2024), protect dopaminergic neurons by enhancing proteasome activity (Haleagrahara and Ponnusamy, 2010) and activate mTOR-S6 signaling to promote local protein synthesis and synaptic density (Gray *et al.*, 2018; Sbrini *et al.*, 2020). These actions address the continuum of age-related diseases by mitigating proteotoxicity and maintaining the integrity of the cellular proteome (Yi *et al.*, 2020).

Collectively, the bioactive constituents of *Centella asiatica*, specifically asiatic acid, asiaticoside, and madecassoside, act as sophisticated modulators of cellular quality control (Figure 1, Table 1). By simultaneously inhibiting harmful protein aggregation and enhancing the clearance mechanisms, such as autophagy and the proteasome, these compounds mitigate the accumulation of unwanted or damaged materials that characterize biological aging. Whether through neuroprotection in the brain or metabolic restoration in the liver and kidneys, *C. asiatica* demonstrates a profound ability to stabilize the proteome, offering a promising therapeutic avenue for delaying the onset of age-related physiological decline and neurodegenerative disease.

3.3. Mitochondrial dysfunction and cellular senescence

Mitochondrial dysfunction and cellular senescence are two fundamental, interconnected drivers of the aging process. Mitochondrial decline is marked by DNA mutations, respiratory complex instability, and impaired mitophagy, further exacerbated by the loss of mitochondrial-derived peptides like humanin and MOTS-c (Reynolds *et al.*, 2021; Somasundaram *et al.*, 2024; Dominiak *et al.*, 2025). This failure in energy production often triggers cellular senescence, a state of permanent proliferative arrest driven by acute or chronic cellular stress (Gorgoulis *et al.*, 2019). Once cells become senescent, they employ a "pro-aging" secretory profile known as the senescence-associated secretory phenotype (SASP), spreading inflammation via cytokines like interleukin-6 (IL-6) and tumor necrosis factor- α (TNF- α), which ultimately leads to chronic conditions like neurodegeneration and atherosclerosis (Birch and Gil, 2020; Xu *et al.*, 2022).

Strategic interventions to preserve mitochondrial integrity focus on optimizing metabolic efficiency and resilience. Key approaches include enhancing fatty acid oxidation via L-carnitine, such as beneficial low-level stress (mitohormesis) with metformin, and plastoquinonyl-decyl-triphenylphosphonium (SkQ1) to neutralize localized oxidative damage (Ratray *et al.*, 2019; Reid; Chee *et al.*, 2021; Thompson *et al.*, 2021). Consequently, safeguarding mitochondrial function has emerged as a primary strategy for extending healthspan and delaying physiological decline. Parallel to mitochondrial maintenance, current anti-aging therapeutics target the accumulation of senescent cells through two distinct pharmacological classes: senolytics, which selectively induce apoptosis in aberrant cells to eliminate them from tissues, and senomorphics, which modulate the cellular phenotype to suppress the SASP without causing cell death. By neutralizing the inflammatory output of these cells, senomorphics mitigate the hallmarks that drives systemic aging.

In cellular models of aging, *Centella asiatica* and its bioactive compounds exhibit pronounced senomorphic effects. Triterpenoids, including asiatic acid, madecassic acid, and madecassoside, attenuate oxidative stress and suppress pro-inflammatory signaling by inhibiting nuclear factor kappa B (NF- κ B). This inhibition effectively reduces the secretion of SASP factors, such as IL-6, IL-1 β , and TNF- α (IL-1 β) (Yun *et al.*, 2008; Dang *et al.*, 2024). Asiaticoside further delays cellular senescence and reduces ROS levels in ultraviolet (UV)-induced

fibroblast models via modulation of the transforming growth factor- β 1/Smad pathway (Jiang et al., 2022). In addition, *C. asiatica* extracts protect dermal fibroblasts against hydrogen peroxide and UVB-induced senescence by modulating microRNA expression (Kim et al., 2011; An et al., 2012).

Beyond cellular systems, these anti-senescence effects are reflected in organismal models. Extracellular vesicles derived from *C. asiatica* promote skin repair in UVB-induced mouse models by enhancing keratinocyte proliferation and collagen deposition (Chang et al., 2025). Furthermore, supplementation with *C. asiatica* has been shown to improve cognitive function while reducing systemic inflammation and oxidative stress markers in human studies (Phoemsapthawee et al., 2022).

At the mitochondrial level, *C. asiatica* exerts context-dependent regulatory effects across different cellular models. In neuronal and oocyte systems, standardized extracts such as ECa 233 and asiatic acid enhance mitochondrial respiration, ATP generation, and membrane potential ($\Delta\Psi_m$). These benefits are mediated through activation of the Nrf2/HO-1 signaling axis, which upregulates essential antioxidant enzymes, including superoxide dismutase (SOD) and glutathione peroxidase (GPx) (Shinomol and Muralidhara, 2008; Xu et al., 2012; Hu et al., 2020).

Whole-organism studies further support these mitochondrial effects. *C. asiatica* attenuates oxidative stress and improve mitochondrial function in brain tissues exposed to neurotoxicity (Boondam et al., 2025). In models of seizure and myocardial ischemia-reperfusion injury, asiatic acid preserves mitochondrial integrity by modulating Bcl-2/Bax balance and restoring key mitochondrial proteins such as sirtuin 3 (SIRT3) and ATP synthase (Lu et al., 2021; Yi et al., 2022). Moreover, in neurodegenerative and metabolic disease models, including amyloid- β toxicity and diabetes, *C. asiatica* alleviates mitochondrial dysfunction by enhancing oxidative respiration and upregulating antioxidant and mitochondrial genes via the Nrf2/HO-1 pathway (Gray et al., 2015; Gray et al., 2018; Giribabu et al., 2020; Kundu et al., 2023).

Overall, *C. asiatica* serves as a dual-action therapeutic agent that stabilizes mitochondrial bioenergetics while simultaneously mitigating the deleterious effects of cellular senescence (Figure 1, Table 1). By upregulating antioxidant defenses via the Nrf2 pathway and suppressing the pro-inflammatory SASP profile, it addresses both the internal metabolic failures and the external inflammatory signaling that characterize aging. This comprehensive ability to restore redox balance and preserve the cellular life cycle positions *C. asiatica* as a significant botanical intervention for extending healthspan and treating age-related pathologies.

3.4. Inflammaging

Inflammaging is defined as a state of chronic, sterile, low-grade systemic inflammation that develops during aging. It is characterized by elevated circulating levels of pro-inflammatory markers, including IL-6, TNF- α , and C-reactive protein (CRP) (Hirata et al., 2020), which collectively contribute to the progression of ARDs such as atherosclerosis and neurodegeneration (Mcgeer & Mcgeer, 2004; López-Otín et al., 2023). This phenomenon arises from a complex interplay of primary aging hallmarks, including genomic instability, the accumulation of the SASP, and impaired autophagy/proteostasis. Furthermore, overactivation of nutrient-sensing pathways, specifically the growth hormone (GH)/insulin-like growth factor 1 (IGF1)/mTORC1 axis, compounded by immune system decline, and gut barrier dysfunction (Miller et al., 2021; Mittelbrunn and Kroemer, 2021; López-Otín et al., 2023). Strategic interventions targeting these inflammatory cascades have demonstrated significant therapeutic potential, showing an ability to reduce systemic inflammation, enhance metabolic and cognitive function, and extend lifespan in various animal models (Gocmez et al., 2020; Marín-Aguilar et al., 2020; Sciorati et al., 2020).

In *in vitro* inflammatory models, the anti-inflammatory effects of *Centella asiatica* and its triterpenoids are typically investigated using lipopolysaccharide (LPS)-induced cell systems.

In these models, macrophages, microglial cells (e.g., BV2), or endometrial epithelial cells are first treated with compounds such as asiatic acid or asiaticoside, followed by LPS stimulation to induce inflammatory responses (Qiu *et al.*, 2015; Cao *et al.*, 2018; Yuyun *et al.*, 2018; Xiao *et al.*, 2025). The activation of key signaling pathways, including NF- κ B and mitogen-activated protein kinase (MAPK), is then evaluated, together with upstream regulators such as interleukin-1 receptor-associated kinase 1 (IRAK1) and transforming growth factor-beta-activated kinase 1 (TAK1), as well as NLR family pyrin domain containing 3 (NLRP3) inflammasome activation (Guo *et al.*, 2015; Cho *et al.*, 2020; He *et al.*, 2024). Specifically, asiatic acid reduces the production of inflammatory mediators, including nitric oxide (NO), prostaglandin E2 (PGE2), and pro-inflammatory cytokines such as TNF- α and IL-6 in LPS-stimulated BV2 microglial cells via modulation of SIRT1 and Notch signaling pathways (Qian *et al.*, 2018; Cho *et al.*, 2020; Md Pizar *et al.*, 2023). Additionally, in disease-related models such as osteoarthritis and psoriasis, treatment with madecassoside or madecassic acid is followed by evaluation of Janus kinase/signal transducer and activator of transcription 3 (JAK/STAT3) and extracellular signal-regulated kinase (ERK)/p38 signaling pathways, along with corresponding changes in inflammatory markers. (Tan *et al.*, 2021; Fu *et al.*, 2022; Lin *et al.*, 2023).

Beyond natural aging models, *C. asiatica* has shown protective effects in models of chronic stress, which is often utilized to simulate accelerated brain aging. In rats subjected to chronic unpredictable stress, *C. asiatica* extract prevented the surge of the pro-inflammatory cytokine TNF- α in the hippocampus. Notably, this anti-inflammatory action occurred independently of BDNF levels, suggesting that the extract targets the 'inflammaging' component of cognitive decline through pathways distinct from neurotrophic support (Ar Rochmah *et al.*, 2019).

These molecular mechanisms provide multi-systemic therapeutic benefits in *in vivo* models. The administration of *C. asiatica* extracts attenuates experimental colitis and dental pulp inflammation by suppressing mitochondrial-mediated inflammatory pathways (Nurhapsari *et al.*, 2023; Shin *et al.*, 2026). In respiratory and renal models, asiaticoside and asiatic acid alleviate acute lung injury and acute kidney injury by reducing inflammatory cell infiltration and stabilizing vascular barrier integrity (Qiu *et al.*, 2015; Sari *et al.*, 2021; Sung *et al.*, 2025; Yang *et al.*, 2018). Furthermore, specialized formulations (Centevita™) also demonstrate significant "anti-inflammaging" and photoprotective effects in murine by activating HO-1 and reducing the accumulation of advanced glycation endproducts (AGEs) (Huang *et al.*, 2011; Lee *et al.*, 2020; Maramaldi *et al.*, 2013; Wan *et al.*, 2013).

Altogether, *C. asiatica* functions as a comprehensive antagonist to inflammaging by targeting the core molecular drivers of systemic inflammation (Figure 1, Table 1). By suppressing the NLRP3 inflammasome and NF- κ B signaling while simultaneously activating protective pathways like HO-1 and SIRT1, it effectively lowers the inflammatory levels of the organism. These findings highlight its therapeutic potential not only in treating acute inflammatory injuries but also in mitigating the chronic, low-grade inflammation that accelerates age-related decline in the brain, skin, and metabolic organs.

3.5. Intercellular communication

Maintaining homeostasis requires the precise coordination of neural, endocrine, and immune signals, a complex regulatory network that inevitably degrades with age. This breakdown is driven by the systemic accumulation of aging factors in the bloodstream, such as CCL11, β 2-microglobulin, IL-6, alongside the chronic secretion of the pro-inflammatory SASP by senescent cells (Smith *et al.*, 2015; Valletta *et al.*, 2020; Villeda *et al.*, 2011). Contemporary strategies to restore this vital communication focus on both neutralizing these harmful circulating elements and harnessing "young" blood factors, such as GDF11, TIMP2, and VEGF. Furthermore, interventions aimed at improving the elasticity of the extracellular matrix, modulating the neuro-endocrine signaling axis, and restoring gut microbiota balance have shown great

promise in rejuvenating immune function. By targeting the systemic environment, these strategies not only preserve the integrity of the cellular communication system but also offer a path toward slowing the aging process and preventing the onset of related multi-systemic diseases.

At the cellular level, *C. asiatica* extracts and their constituents actively orchestrate cellular differentiation and tissue repair. Human mesenchymal stem cells (hMSCs) and neuro-differentiated mesenchymal stem cells (ndMSCs) conditioned with these extracts undergo accelerated neural differentiation, characterized by enhanced axonal regeneration, increased myelin thickness, and optimized synaptic connectivity (Hussin et al., 2020; Omar et al., 2019). Within the intricate framework of the central nervous system, water extracts and their caffeoylquinic acid constituents stimulate mouse primary hippocampal neurons to expand their dendritic arborization and synaptic complexity (Rowe et al., 2024). Furthermore, asiatic acid derivatives and araliadiol modulate the extracellular matrix by upregulating osteogenic proteins (BMP2, WNT3A, ALP) via ERK signaling and promoting hair growth through the p38/PPAR- γ axis (Park et al., 2024; Thanmmium et al., 2023).

Translating these cellular findings into a systemic context, *in vivo* research demonstrates significant functional recovery. The administration of water extracts effectively mitigates age-related locomotion deficits in models such as *Drosophila* by preserving neuronal arborization (Rowe et al., 2024). Moreover, the transplantation of ndMSCs pre-treated with *C. asiatica* significantly accelerates the physical repair of peripheral nerves, restoring both structural integrity and nerve conduction velocity in injury models (Hussin et al., 2020). These results suggest that the botanical not only provides the raw signaling cues for repair but also enhances the functional capacity of transplanted cells to integrate into damaged tissues.

In conclusion, *C. asiatica* supports intercellular communication during aging by enhancing synaptic connectivity and regulating ECM signaling pathways (Figure 1, Table 1). By fostering a microenvironment conducive to stem cell differentiation and axonal growth, it addresses the age-related decline in tissue regenerative capacity. This ability to modulate the structural and chemical dialogue between cells, whether through the upregulation of osteogenic proteins or the preservation of neuronal architecture, positions *C. asiatica* as a vital agent for maintaining systemic coordination and functional integrity throughout the aging process.

3.6. Antioxidant mechanisms

Under physiological conditions, ROS act as critical signaling molecules. However, during aging, the progressive decline in mitochondrial efficiency driven by mtDNA mutations and proteostatic imbalance leads to a pathological surge in ROS production (Amorim et al., 2022; López-Otín & Kroemer, 2021). When the leakage of ROS or mtDNA into the cytoplasm exceeds the cell's neutralizing capacity, it triggers a cascade of genetic damage, lipid peroxidation, and accelerated telomere shortening (Amorim et al., 2022; López-Otín & Kroemer, 2021). To mitigate this progression, antioxidants play a central role in preserving genomic integrity, stabilizing cellular membranes, and maintaining mitochondrial function while suppressing the chronic inflammatory responses associated with the aging process.

A primary mechanism for counteracting this cellular toxicity is the direct scavenging of ROS, which prevents their deleterious interaction with macromolecules like DNA. Non-enzymatic antioxidants, such as vitamin C, vitamin E, glutathione, and natural polyphenols, neutralize ROS by donating electrons or hydrogen atoms, converting them into stable, non-oxidizing molecules. Complementing this direct action, the body's enzymatic antioxidant system comprising SOD, catalase (CAT), and GPx actively metabolizes and eliminates free radicals (Hossain et al., 2022; Petr et al., 2020). Together, these systems preserve the structural and functional integrity of the cell against the oxidative pressures that define biological aging.

Beyond direct scavenging by vitamins and polyphenols, *C. asiatica* provides advanced cellular defense by activating of the Nrf2/antioxidant response element (ARE) pathway (Kim & Jeon, 2022). In *in vitro* settings, its constituents promote the dissociation of Nrf2 from its

negative regulator, Kelch-like ECH-associated protein 1 (Keap1), thereby inducing the expression of cytoprotective enzymes. These include heme oxygenase-1 (HO-1), NAD(P)H quinone dehydrogenase 1 (NQO1), glutamate-cysteine ligase catalytic subunit (GCLC), SOD, CAT, and GPx (Papaiahgari et al., 2006; Qi et al., 2017). Specifically, asiatic acid activates Nrf2 via protein kinase B (AKT) and ERK signaling to bolster cellular resilience (Qi et al., 2017). Interestingly, while ethanolic extracts provide broad cytoprotection, specific fractions can modulate differentially in contexts. Madecassoside preserves mitochondrial membrane potential ($\Delta\Psi_m$) and stimulates autophagy to clear damaged organelles in human umbilical vein endothelial cells (HUVECs) and melanocytes, protecting them against oxidative stress-induced injury (Bian et al., 2012; Ling et al., 2017).

Translating to *in vivo* models, these mechanisms provide robust protection across multiple systems. In skin aging and photoaging, *C. asiatica* and its extracellular vesicles (EVs) attenuate hydrogen peroxide-induced damage, inhibit collagenase and elastase, and restore cutaneous density and hydration (Firdaus et al., 2024; Haftek et al., 2008; Kim et al., 2023; Zofia et al., 2020). These findings are supported by clinical evaluations of topical formulations, such as water-in-oil creams containing *C. asiatica* and ferulic acid, which demonstrate significant increases in cutaneous density, hydration, and overall antioxidant activity (Moldovan et al., 2017; Ramesh et al., 2014).

In neurodegenerative models, including Parkinson's disease (PD) and Alzheimer's disease (AD), asiatic acid acts as a mitochondrial complex-I protector, preserving $\Delta\Psi_m$ and reducing amyloid- β -induced oxidative damage (Gray et al., 2017; Nataraj et al., 2017; Rather et al., 2018). These effects are further supported by the restoration of cholinergic function and enhancement of electron transport chain activity, effectively mitigating cognitive impairment and age-related neurological decline (Chiroma et al., 2019; Kumar et al., 2011; Mansor et al., 2023; Subathra et al., 2005; Zhong et al., 2025). Additionally, these antioxidant benefits extend to reproductive health, where asiatic acid improves reproductive competence by enhancing ATP production and blastocyst development under oxidative stress (Qi et al., 2021).

In conclusion, *Centella asiatica* provides a sophisticated, multi-level antioxidant defense that transcends simple free-radical scavenging (Figure 1, Table 1). By orchestrating the genetic response and safeguarding mitochondrial complex activity, it reinforces the cell's internal machinery against the primary drivers of aging. Whether through protecting the structural integrity of the skin, preserving cognitive function in the brain, or stabilizing systemic metabolic health, *C. asiatica* demonstrates a unique ability to mitigate aging-related damage across diverse tissues, making it a powerful candidate for geroscience-based interventions.

3.7 Beyond the core hallmarks

In addition to addressing the core molecular hallmarks of aging, *C. asiatica* modulates systemic physiology and complex behaviors, offering a holistic impact on the aging organism (Figure 2).

High-dose water extracts of *C. asiatica* (1000 mg/kg/day) have been reported to improve REM sleep, enhance cognitive performance, and reduce anxiety-like behaviors in aging and Alzheimer's animal models (Gray et al., 2024; Kitzes et al., 1985; Ridker et al., 2017; Speers et al., 2024). A notable neuroprotective mechanism was recently identified in the standardized extract ECa 233, which demonstrated a significant reduction in brain iron accumulation (Yatmark et al., 2025).

In the clinical sphere, *C. asiatica* has established a strong presence in cosmeceutical applications. Topical formulations have been shown to reduce wrinkle depth and improve skin hydration, two primary markers of dermatological aging commonly observed in the elderly (Kongkaew et al., 2020). Advanced formulations, such as those combining the plant with mandelic acid, further enhance skin thickness and collagen synthesis while reinforcing barrier integrity to promote tissue repair (Widgerow et al., 2025). Although many of these clinical trials

are relatively small, they provide moderate, evidence-based support for its efficacy in dermatological anti-aging.



Figure 2. Multi-system protective effects and biological activities of *Centella asiatica*. The radius of each pie chart segment corresponds to the strength of evidence from direct research.

C. asiatica demonstrates robust neuroprotective and cognitive benefits, followed by significant effects on skin repair, collagen synthesis, and protection against UV-induced damage.

Additionally, it supports cardiovascular function and overall systemic health. Created in Adobe Illustrator.

Mechanistic insights have further highlighted the role of triterpenes and caffeoylquinic acids in regulating genes involved in extracellular matrix organization, metabolism, and immune signalling. Such molecular actions likely contribute to the improvements in cognitive and behavioural functions observed in animal models (Chamberlin et al., 2024; Hack et al., 2025; Tsoukalas et al., 2021). These findings emphasize that *C. asiatica* acts not only at the cellular and molecular levels but also influences systemic physiology and inter-organ interactions.

Taken together, *C. asiatica* presents a promising profile of neuroprotective, and cosmeceutical properties that extend its utility beyond basic antioxidant support. While clinical evidence is most robust in dermatological applications, its potential to stabilize sleep patterns, reduce and neurotoxic iron loading provides a compelling case for its role as a systemic anti-aging agent.

Discussion

Centella asiatica exhibits a diverse range of bioactivities that directly address the core drivers of aging, primarily through modulation of oxidative stress, mitochondrial function, and inflammation. Its major triterpenoids, including asiaticoside, madecassoside, asiatic acid, and madecassic acid, have demonstrated potent antioxidant and cytoprotective effects across multiple experimental systems. These compounds work by reducing intracellular ROS, preserving mitochondrial integrity, and regulating apoptosis-related signalling (Ling et al., 2017). These molecular actions are further supported by *in vivo* studies highlighting neuroprotective and cardioprotective benefits (Nataraj et al., 2017; Qiu et al., 2022). Importantly, the efficacy

of *C. asiatica* lies in its ability to target interconnected biological processes central to the “Hallmarks of Aging” (López-Otín *et al.*, 2013), suggesting that its constituents act on integrated mechanistic nodes rather than isolated targets. This interconnectedness suggests that these pathways may represent key mechanistic nodes underlying the potential anti-aging effects of *C. asiatica*, rather than acting as isolated targets.

Despite this promising evidence, several factors limit the current literature. Most studies are confined to *in vitro* systems or animal models, which often fail to replicate the complexity of human physiology and systemic pharmacokinetic interactions (Gray *et al.*, 2017; Lu *et al.*, 2016). A critical challenge is the limited bioavailability of triterpenoid compounds. Due to poor absorption and rapid metabolism, oral administration often results in low systemic exposure (He *et al.*, 2023; Yuan *et al.*, 2015). Moreover, many *in vitro* studies utilize micromolar concentrations that exceed physiologically achievable levels *in vivo*, creating a significant gap in extrapolating experimental efficacy to clinical outcomes (Songvut *et al.*, 2021; Tan *et al.*, 2021). These discrepancies highlight a key limitation in extrapolating *in vitro* findings to *in vivo* contexts and may partially explain inconsistencies between experimental efficacy and clinical outcomes. In addition, variations in plant sources and extraction methods lead to phytochemical heterogeneity, complicating reproducibility (Idris and Mohd Nadzir, 2021). Furthermore, long-term safety data and potential herb-drug interactions remain under-researched (Hein *et al.*, 2025).

Future research should address these limitations through strategically designed studies aimed at enhancing translational applicability. Priority should be given to randomized, placebo-controlled clinical trials utilizing standardized extracts with well-characterized phytochemical profiles and endpoints related to aging hallmarks. Parallel investigations on pharmacokinetics and formulation development are essential to improve bioavailability and establish robust dose-response relationships. Long-term safety studies across diverse populations are also necessary to ensure generalizability and clinical relevance. Addressing these gaps will be crucial for bridging the current divide between preclinical findings and therapeutic implementation.

In conclusion, *C. asiatica* represents a promising multi-target botanical with significant anti-aging potential. While further clinical validation is required, current preclinical evidence highlights its potential to contribute to strategies aimed at delaying aging and improving healthspan.

Authors' contributions:

Research concept and design, Nguyen Ngoc NA, Pham Thi TT, Ngo Huynh BC, Do Thi AT, Ly Hong HH; Collection and assembly of data, data analysis and interpretation, and writing the article, Nguyen Ngoc NA, Pham Thi TT, Ngo Huynh BC, collection of data, Tran AV; Critical revision of the article, Do Thi AT, Ly Hong HH; and Final approval of the article, Ly Hong HH. Nguyen Ngoc NA, Pham Thi TT, and Ngo Huynh BC contributed equally to the study.

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Conflict of interests

The authors declare that they have no conflicts of interest that could have influenced the conduct, analysis or presentation of the research.

References

- Alsaleh G, Panse I, Swadling L, Zhang H, Richter FC, Meyer A, Lord J, Barnes E, Klenerman P, Green C & Simon AK (2020) Autophagy in T cells from aged donors is maintained by spermidine and correlates with function and vaccine responses. *Elife* 9:DOI: 10.7554/eLife.57950.
- Amorim JA, Coppotelli G, Rolo AP, Palmeira CM, Ross JM & Sinclair DA (2022) Mitochondrial and metabolic dysfunction in ageing and age-related diseases. *Nat Rev Endocrinol* 18:243-58 DOI: 10.1038/s41574-021-00626-7.
- An I-S, An S, Kang S-M, Choe T-B, Lee SN, Jang HH & Bae S (2012) Titrated extract of *Centella asiatica* provides a UVB protective effect by altering microRNA expression profiles in human dermal fibroblasts. *Int J Mol Med* 30:1194-202 DOI: 10.3892/ijmm.2012.1117.
- Ar Rochmah M, Harini IM, Septyaningtrias DE, Sari DCR & Susilowati R (2019) *Centella asiatica* Prevents Increase of Hippocampal Tumor Necrosis Factor- α Independently of Its Effect on Brain-Derived Neurotrophic Factor in Rat Model of Chronic Stress. *Biomed Res Int* 2019:2649281 DOI: 10.1155/2019/2649281.
- Bian D, Liu M, Li Y, Xia Y, Gong Z & Dai Y (2012) Madecassoside, a triterpenoid saponin isolated from *Centella asiatica* herbs, protects endothelial cells against oxidative stress. *Journal of Biochemical and Molecular Toxicology* 26:399-406 DOI: <https://doi.org/10.1002/jbt.21434>.
- Birch J & Gil J (2020) Senescence and the SASP: many therapeutic avenues. *Genes Dev* 34:1565-76 DOI: 10.1101/gad.343129.120.
- Blackburn EH, Epel ES & Lin J (2015) Human telomere biology: A contributory and interactive factor in aging, disease risks, and protection. *Science* 350:1193-8 DOI: doi:10.1126/science.aab3389.
- Blasco MA (2005) Telomeres and human disease: ageing, cancer and beyond. *Nature Reviews Genetics* 6:611-22 DOI: 10.1038/nrg1656.
- Boondam Y, Pakaprot N, Yang MC, Sandech N, Maiuthed A, Samer J, Prasittisa K, Ruanpang J, Care C & Chuayboon S (2025) Comparative effects of standardized *Centella asiatica* extract (ECa 233) and its active compound mixture on proteomics and mitochondrial function. *Scientific Reports* 15:29348 DOI: 10.1038/s41598-025-12622-2.
- Cao S-y, Wang W, Nan F-f, Liu Y-n, Wei S-y, Li F-f & Chen L (2018) Asiatic acid inhibits LPS-induced inflammatory response in endometrial epithelial cells. *Microbial Pathogenesis* 116:195-9 DOI: <https://doi.org/10.1016/j.micpath.2018.01.022>.
- Chakravarti D, LaBella KA & DePinho RA (2021) Telomeres: history, health, and hallmarks of aging. *Cell* 184:306-22 DOI: 10.1016/j.cell.2020.12.028.
- Chamberlin S, Zweig JA, Neff CJ, Marney L, Choi J, Yang L, Maier CS, Soumyanath A, McWeeney S & Gray NE (2024) Multi-omics analysis in mouse primary cortical neurons reveals complex positive and negative biological interactions between constituent compounds in *Centella asiatica*. *bioRxiv* DOI: 10.1101/2024.11.04.621595.
- Chang TM, Wu CC, Huang HC, Wang SS, Chuang CH, Kao PL, Tang WH, Liu LT, Qiu WY, Percec I, Chen C & Kuo TY (2025) In Vitro Characterization of *Centella asiatica* Extracellular Vesicles and Their Skin Repair Effects in a UVB-Irradiated Mouse Model. *Int J Mol Sci* 26:DOI: 10.3390/ijms26188982.
- Chee C, Shannon CE, Burns A, Selby AL, Wilkinson D, Smith K, Greenhaff PL & Stephens FB (2021) Increasing skeletal muscle carnitine content in older individuals increases whole-body fat oxidation during moderate-intensity exercise. *Aging Cell* 20:e13303 DOI: 10.1111/acel.13303.
- Chiroma SM, Hidayat Baharuldin MT, Mat Taib CN, Amom Z, Jagadeesan S, Adenan MI & Mohd Moklas MA (2019) Protective effect of *Centella asiatica* against D-galactose and aluminium chloride induced rats: Behavioral and ultrastructural approaches.

- Cho Y-C, Vuong HL, Ha J, Lee S, Park J, Wibow AE & Cho S (2020) Inhibition of Inflammatory Responses by Centella asiatica via Suppression of IRAK1-TAK1 in Mouse Macrophages. *The American Journal of Chinese Medicine* 48:1103-20 DOI: 10.1142/s0192415x20500548.
- Choi SW, Cho W, Oh H, Abd El-Aty AM, Hong SA, Hong M, Jeong JH & Jung TW (2023) Madecassoside ameliorates hepatic steatosis in high-fat diet-fed mice through AMPK/autophagy-mediated suppression of ER stress. *Biochemical Pharmacology* 217:115815 DOI: <https://doi.org/10.1016/j.bcp.2023.115815>.
- Dang Y-y, Liu T, Liu Y-d, Li J-y, Jing Y, Yang M-j, Zhang H, Jiang M-m, Wu H-h, Yang W-z, Li N & Zhang P (2024) Anti-photoaging activity of triterpenoids isolated from Centella asiatica. *Phytochemistry* 228:114246 DOI: <https://doi.org/10.1016/j.phytochem.2024.114246>.
- Demidenko O, Barardo D, Budovskii V, Fimmemore R, Palmer FR, Kennedy BK & Budovskaya YV (2021) Rejuvant®, a potential life-extending compound formulation with alpha-ketoglutarate and vitamins, conferred an average 8 year reduction in biological aging, after an average of 7 months of use, in the TruAge DNA methylation test. *Ageing (Albany NY)* 13:24485-99 DOI: 10.18632/aging.203736.
- Dhanasekaran M, Holcomb LA, Hitt AR, Tharakan B, Porter JW, Young KA & Manyam BV (2009) Centella asiatica extract selectively decreases amyloid beta levels in hippocampus of Alzheimer's disease animal model. *Phytother Res* 23:14-9 DOI: 10.1002/ptr.2405.
- Dominiak A, Gawinek E, Banaszek AA & Wilkaniec A (2025) Mitochondrial Quality Control in Neurodegeneration and Cancer: A Common Denominator, Distinct Therapeutic Challenges. *International Journal of Molecular Sciences* 26:8693.
- Firdaus Z, Gutti G, Ganeshpurkar A, Kumar A, Krishnamurthy S, Singh SK & Singh TD (2024) Centella asiatica improves memory and executive function in middle-aged rats by controlling oxidative stress and cholinergic transmission. *Journal of Ethnopharmacology* 325:117888 DOI: <https://doi.org/10.1016/j.jep.2024.117888>.
- Franceschi C, Garagnani P, Morsiani C, Conte M, Santoro A, Grignolio A, Monti D, Capri M & Salvioli S (2018) The Continuum of Aging and Age-Related Diseases: Common Mechanisms but Different Rates. *Frontiers in Medicine* Volume 5 - 2018:DOI: 10.3389/fmed.2018.00061.
- Fu X, He S, Wang L, Xue Y, Qiao S, An J & Xia T (2022) Madecassic Acid Ameliorates the Progression of Osteoarthritis: An in vitro and in vivo Study. *Drug Des Devel Ther* 16:3793-804 DOI: 10.2147/dddt.S383632.
- Gerashchenko MV, Peterfi Z, Yim SH & Gladyshev VN (2021) Translation elongation rate varies among organs and decreases with age. *Nucleic Acids Res* 49:e9 DOI: 10.1093/nar/gkaa1103.
- Gomez SS, Yazir Y, Gacar G, Demirtaş Şahin T, Arkan S, Karson A & Utkan T (2020) Etanercept improves aging-induced cognitive deficits by reducing inflammation and vascular dysfunction in rats. *Physiology & Behavior* 224:113019 DOI: <https://doi.org/10.1016/j.physbeh.2020.113019>.
- Gorbunova V, Seluanov A, Mita P, McKerrow W, Fenyö D, Boeke JD, Linker SB, Gage FH, Kreiling JA, Petrashen AP, Woodham TA, Taylor JR, Helfand SL & Sedivy JM (2021) The role of retrotransposable elements in ageing and age-associated diseases. *Nature* 596:43-53 DOI: 10.1038/s41586-021-03542-y.
- Gorgoulis V, Adams PD, Alimonti A, Bennett DC, Bischof O, Bishop C, Campisi J, Collado M, Evangelou K, Ferbeyre G, Gil J, Hara E, Krizhanovsky V, Jurk D, Maier AB, Narita M, Niedernhofer L, Passos JF, Robbins PD, Schmitt CA, Sedivy J, Vougas K, von

- Zglinicki T, Zhou D, Serrano M & Demaria M (2019) Cellular Senescence: Defining a Path Forward. *Cell* 179:813-27 DOI: 10.1016/j.cell.2019.10.005.
- Gray NE, Hack W, Brandes MS, Zweig JA, Yang L, Marney L, Choi J, Magana AA, Cerruti N, McFerrin J, Koike S, Nguyen T, Raber J, Quinn JF, Maier CS & Soumyanath A (2024) Amelioration of age-related cognitive decline and anxiety in mice by *Centella asiatica* extract varies by sex, dose and mode of administration. *Front Aging* 5:1357922 DOI: 10.3389/fragi.2024.1357922.
- Gray NE, Sampath H, Zweig JA, Quinn JF & Soumyanath A (2015) *Centella asiatica* Attenuates Amyloid- β -Induced Oxidative Stress and Mitochondrial Dysfunction. *J Alzheimers Dis* 45:933-46 DOI: 10.3233/jad-142217.
- Gray NE, Zweig JA, Caruso M, Martin MD, Zhu JY, Quinn JF & Soumyanath A (2018) *Centella asiatica* increases hippocampal synaptic density and improves memory and executive function in aged mice. *Brain Behav* 8:e01024 DOI: 10.1002/brb3.1024.
- Gray NE, Zweig JA, Caruso M, Zhu JY, Wright KM, Quinn JF & Soumyanath A (2018) *Centella asiatica* attenuates hippocampal mitochondrial dysfunction and improves memory and executive function in β -amyloid overexpressing mice. *Mol Cell Neurosci* 93:1-9 DOI: 10.1016/j.mcn.2018.09.002.
- Gray NE, Zweig JA, Matthews DG, Caruso M, Quinn JF & Soumyanath A (2017) *Centella asiatica* Attenuates Mitochondrial Dysfunction and Oxidative Stress in A β -Exposed Hippocampal Neurons. *Oxid Med Cell Longev* 2017:7023091 DOI: 10.1155/2017/7023091.
- Guo W, Liu W, Jin B, Geng J, Li J, Ding H, Wu X, Xu Q, Sun Y & Gao J (2015) Asiatic acid ameliorates dextran sulfate sodium-induced murine experimental colitis via suppressing mitochondria-mediated NLRP3 inflammasome activation. *International Immunopharmacology* 24:232-8 DOI: <https://doi.org/10.1016/j.intimp.2014.12.009>.
- Giribabu N, Karim K, Kilari EK, Nelli SR & Salleh N (2020) Oral administration of *Centella asiatica* (L.) Urb leave aqueous extract ameliorates cerebral oxidative stress, inflammation, and apoptosis in male rats with type-2 diabetes. *Inflammopharmacology* 28:1599-622 DOI: 10.1007/s10787-020-00733-3.
- Hack W, Kuhnu L, Martinez J, Marney LC, Choi J, Sohal AR, Koike S, Nguyen T, Maier CS, Soumyanath A & Gray NE (2025) Triterpene and Caffeoylquinic Acid Constituents Contribute to the Cognitive-Enhancing, but Not Anxiolytic, Effects of a Water Extract of *Centella asiatica* in Aged Mice. *Nutrients* 17:DOI: 10.3390/nu17193171.
- Haftik M, Mac-Mary S, Bitoux M-AL, Creidi P, Seité S, Rougier A & Humbert P (2008) Clinical, biometric and structural evaluation of the long-term effects of a topical treatment with ascorbic acid and madecassoside in photoaged human skin. *Experimental Dermatology* 17:946-52 DOI: <https://doi.org/10.1111/j.1600-0625.2008.00732.x>.
- Haleagrahara N & Ponnusamy K (2010) Neuroprotective effect of *Centella asiatica* extract (CAE) on experimentally induced parkinsonism in aged Sprague-Dawley rats. *The Journal of Toxicological Sciences* 35:41-7 DOI: 10.2131/jts.35.41.
- Hao Y, Huang J, Ma Y, Chen W, Fan Q, Sun X, Shao M & Cai H (2018) Asiatic acid inhibits proliferation, migration and induces apoptosis by regulating Pcd4 via the PI3K/Akt/mTOR/p70S6K signaling pathway in human colon carcinoma cells. *Oncol Lett* 15:8223-30 DOI: 10.3892/ol.2018.8417.
- He Z, Hu Y, Niu Z, Zhong K, Liu T, Yang M, Ji L & Hu W (2023) A review of pharmacokinetic and pharmacological properties of asiaticoside, a major active constituent of *Centella asiatica* (L.) Urb. *J Ethnopharmacol* 302:115865 DOI: 10.1016/j.jep.2022.115865.
- He Z, Hu Y, Zhang Y, Xie J, Niu Z, Yang G, Zhang J, Zhao Z, Wei S, Wu H & Hu W (2024) Asiaticoside exerts neuroprotection through targeting NLRP3 inflammasome activation. *Phytomedicine* 127:155494 DOI: <https://doi.org/10.1016/j.phymed.2024.155494>.

- Hein ZM, Gopalakrishna PK, Kanuri AK, Thomas W, Hussan F, Naik VR, Shantakumari N, Che Ramli MD, Mohd Moklas MA, Che Mohd Nassir CMN & Vishnumukkala T (2025) *Centella asiatica*: Advances in Extraction Technologies, Phytochemistry, and Therapeutic Applications. *Life* 15:1081.
- Hetz C, Zhang K & Kaufman RJ (2020) Mechanisms, regulation and functions of the unfolded protein response. *Nat Rev Mol Cell Biol* 21:421-38 DOI: 10.1038/s41580-020-0250-z.
- Hipp MS, Kasturi P & Hartl FU (2019) The proteostasis network and its decline in ageing. *Nat Rev Mol Cell Biol* 20:421-35 DOI: 10.1038/s41580-019-0101-y.
- Hirata T, Arai Y, Yuasa S, Abe Y, Takayama M, Sasaki T, Kunitomi A, Inagaki H, Endo M, Morinaga J, Yoshimura K, Adachi T, Oike Y, Takebayashi T, Okano H & Hirose N (2020) Associations of cardiovascular biomarkers and plasma albumin with exceptional survival to the highest ages. *Nat Commun* 11:3820 DOI: 10.1038/s41467-020-17636-0.
- Hossain R, Quispe C, Khan RA, Saikat ASM, Ray P, Ongalbek D, Yeskaliyeva B, Jain D, Smeriglio A, Trombetta D, Kiani R, Kobarfard F, Mojgani N, Saffarian P, Ayatollahi SA, Sarkar C, Islam MT, Keriman D, Uçar A, Martorell M, Sureda A, Pintus G, Butnariu M, Sharifi-Rad J & Cho WC (2022) Propolis: An update on its chemistry and pharmacological applications. *Chin Med* 17:100 DOI: 10.1186/s13020-022-00651-2.
- Hou W-S, Luo Y-H, Wu N, Tang Y-J, Liu Y-Z, Zhang Y-L & Jin C-H (2025) Madecassoside Induces Apoptosis and Inhibits Migration by Regulating ROS-Mediated Signaling Pathways in MDA-MB-231 Breast Cancer Cells. *Chemical Biology & Drug Design* 106:e70197 DOI: <https://doi.org/10.1111/cbdd.70197>.
- Hu WY, Li XX, Diao YF, Qi JJ, Wang DL, Zhang JB, Sun BX & Liang S (2020) Asiatic acid protects oocytes against in vitro aging-induced deterioration and improves subsequent embryonic development in pigs. *Ageing (Albany NY)* 13:3353-67 DOI: 10.18632/aging.202184.
- Huang SS, Chiu CS, Chen HJ, Hou WC, Sheu MJ, Lin YC, Shie PH & Huang GJ (2011) Antinociceptive activities and the mechanisms of anti-inflammation of asiatic Acid in mice. *Evid Based Complement Alternat Med* 2011:895857 DOI: 10.1155/2011/895857.
- Hussin HM, Lawi MM, Haflah NHM, Kassim AYM, Idrus RBH & Lokanathan Y (2020) *Centella asiatica* (L.)-Neurodifferentiated Mesenchymal Stem Cells Promote the Regeneration of Peripheral Nerve. *Tissue Eng Regen Med* 17:237-51 DOI: 10.1007/s13770-019-00235-6.
- Ibrahim N, Nadian I, Noor DR & Fadilah F (2023) Prediction of Translational Regulation by Network Interaction in Synaptic Plasticity Induced with *Centella asiatica*. *ScientificWorldJournal* 2023:4199614 DOI: 10.1155/2023/4199614.
- Idris FN & Mohd Nadzir M (2021) Comparative Studies on Different Extraction Methods of *Centella asiatica* and Extracts Bioactive Compounds Effects on Antimicrobial Activities. *Antibiotics* 10:457.
- James JT & Dubery IA (2009) Pentacyclic triterpenoids from the medicinal herb, *Centella asiatica* (L.) Urban. *Molecules* 14:3922-41 DOI: 10.3390/molecules14103922.
- Jaskelioff M, Muller FL, Paik JH, Thomas E, Jiang S, Adams AC, Sahin E, Kost-Alimova M, Protopopov A, Cadiñanos J, Horner JW, Maratos-Flier E & Depinho RA (2011) Telomerase reactivation reverses tissue degeneration in aged telomerase-deficient mice. *Nature* 469:102-6 DOI: 10.1038/nature09603.
- Jiang H, Zhou X & Chen L (2022) Asiaticoside delays senescence and attenuate generation of ROS in UV-exposure cells through regulates TGF- β 1/Smad pathway. *Exp Ther Med* 24:667 DOI: 10.3892/etm.2022.11603.
- Joy J, Alarifi S, Alsuhaibani E & Nair CKK (2015) Protection of DNA From Ionizing Radiation-Induced Lesions by Asiaticoside. 34:353-61 DOI: 10.1615/JEnvironPatholToxicolOncol.2015013946.

- Karsono AH, Tandrasasmita OM, Berlian G & Tjandrawinata RR (2021) Potential Antiaging Effects of DLBS1649, a *Centella asiatica* Bioactive Extract. *J Exp Pharmacol* 13:781-95 DOI: 10.2147/jep.S299547.
- Kim MJ & Jeon JH (2022) Recent Advances in Understanding Nrf2 Agonism and Its Potential Clinical Application to Metabolic and Inflammatory Diseases. *Int J Mol Sci* 23:DOI: 10.3390/ijms23052846.
- Kim MJ, Ko H, Kim JY, Kim HJ, Kim HY, Cho HE, Cho HD, Seo WS & Kang HC (2023) Improvement in Yield of Extracellular Vesicles Derived from Edelweiss Callus Treated with LED Light and Enhancement of Skin Anti-Aging Indicators. *Curr Issues Mol Biol* 45:10159-78 DOI: 10.3390/cimb45120634.
- Kim YJ, Cha HJ, Nam KH, Yoon Y, Lee H & An S (2011) *Centella asiatica* extracts modulate hydrogen peroxide-induced senescence in human dermal fibroblasts. *Experimental Dermatology* 20:998-1003 DOI: <https://doi.org/10.1111/j.1600-0625.2011.01388.x>.
- Kitzes R, Ackerman Z, Levy M, Frummerman I, Shved A & Garty M (1985) Bioavailability of phenytoin: comparison between three preparations. *Isr J Med Sci* 21:323-6.
- Kongkaew C, Meesomperm P, Scholfield CN, Chaiwiang N & Waranuch N (2020) Efficacy and Safety of *Centella Asiatica* (L.) Urb. on Wrinkles: A Systematic Review of Published Data and Network Meta-Analysis. *J Cosmet Sci* 71:439-54.
- Kumar A, Prakash A & Dogra S (2011) *Centella asiatica* Attenuates D-Galactose-Induced Cognitive Impairment, Oxidative and Mitochondrial Dysfunction in Mice. *Int J Alzheimers Dis* 2011:347569 DOI: 10.4061/2011/347569.
- Kundu P, Yasuhara K, Brandes MS, Zweig JA, Neff CJ, Holden S, Kessler K, Matsumoto S, Offner H, Waslo CS, Vandenbark A, Soumyanath A, Sherman LS, Raber J, Gray NE & Spain RI (2023) *Centella asiatica* promotes antioxidant gene expression and mitochondrial oxidative respiration in experimental autoimmune encephalomyelitis. *Res Sq* DOI: 10.21203/rs.3.rs-3393042/v1.
- Lee Y, Choi HK, N'Deh K PU, Choi YJ, Fan M, Kim EK, Chung KH & An AJH (2020) Inhibitory Effect of *Centella asiatica* Extract on DNCB-Induced Atopic Dermatitis in HaCaT Cells and BALB/c Mice. *Nutrients* 12:DOI: 10.3390/nu12020411.
- Li C, Meng X, Li S, Wu Q & Wang C (2025) Madecassoside Accelerates Nerve Regeneration by Promoting M2 Macrophage Polarization via TXNIP/NLRP3/GSDMD Pathway. *Molecular Neurobiology* 62:15687-700 DOI: 10.1007/s12035-025-05196-7.
- Limanaqi F, Biagioni F, Busceti CL, Ryskalin L, Polzella M, Frati A & Fornai F (2019) Phytochemicals Bridging Autophagy Induction and Alpha-Synuclein Degradation in Parkinsonism. *Int J Mol Sci* 20:DOI: 10.3390/ijms20133274.
- Lin P, Shi H-y, Lu Y-y & Lin J (2023) *Centella asiatica* alleviates psoriasis through JAK/STAT3-mediated inflammation: An in vitro and in vivo study. *Journal of Ethnopharmacology* 317:116746 DOI: <https://doi.org/10.1016/j.jep.2023.116746>.
- Ling Y, Gong Q, Xiong X, Sun L, Zhao W, Zhu W & Lu Y (2017) Protective effect of madecassoside on H₂O₂-induced oxidative stress and autophagy activation in human melanocytes. *Oncotarget* 8:51066-75 DOI: 10.18632/oncotarget.17654.
- Ling Z, Zhou S, Zhou Y, Zhong W, Su Z & Qin Z (2024) Protective role of madecassoside from *Centella asiatica* against protein L-isoaspartyl methyltransferase deficiency-induced neurodegeneration. *Neuropharmacology* 246:109834 DOI: <https://doi.org/10.1016/j.neuropharm.2023.109834>.
- Lipinski MM, Zheng B, Lu T, Yan Z, Py BF, Ng A, Xavier RJ, Li C, Yankner BA, Scherzer CR & Yuan J (2010) Genome-wide analysis reveals mechanisms modulating autophagy in normal brain aging and in Alzheimer's disease. *Proc Natl Acad Sci U S A* 107:14164-9 DOI: 10.1073/pnas.1009485107.
- Liu R, Wu CX, Zhou D, Yang F, Tian S, Zhang L, Zhang TT & Du GH (2012) Pinocembrin protects against β -amyloid-induced toxicity in neurons through inhibiting receptor for

- advanced glycation end products (RAGE)-independent signaling pathways and regulating mitochondrion-mediated apoptosis. *BMC Med* 10:105 DOI: 10.1186/1741-7015-10-105.
- López-Otín C, Blasco MA, Partridge L, Serrano M & Kroemer G (2013) The hallmarks of aging. *Cell* 153:1194-217 DOI: 10.1016/j.cell.2013.05.039.
- López-Otín C, Blasco MA, Partridge L, Serrano M & Kroemer G (2023) Hallmarks of aging: An expanding universe. *Cell* 186:243-78 DOI: 10.1016/j.cell.2022.11.001.
- López-Otín C & Kroemer G (2021) Hallmarks of Health. *Cell* 184:33-63 DOI: 10.1016/j.cell.2020.11.034.
- López-Otín C, Pietrocola F, Roiz-Valle D, Galluzzi L & Kroemer G (2023) Meta-hallmarks of aging and cancer. *Cell Metabolism* 35:12-35 DOI: 10.1016/j.cmet.2022.11.001.
- Lu CW, Lin TY, Pan TL, Wang PW, Chiu KM, Lee MY & Wang SJ (2021) Asiatic Acid Prevents Cognitive Deficits by Inhibiting Calpain Activation and Preserving Synaptic and Mitochondrial Function in Rats with Kainic Acid-Induced Seizure. *Biomedicines* 9:DOI: 10.3390/biomedicines9030284.
- Lu Y, Liu S, Wang Y, Wang D, Gao J & Zhu L (2016) Asiatic acid uncouples respiration in isolated mouse liver mitochondria and induces HepG2 cells death. *European Journal of Pharmacology* 786:212-23 DOI: <https://doi.org/10.1016/j.ejphar.2016.06.010>.
- Luo P, Huang Q, Chen S, Wang Y & Dou H (2022) Asiaticoside ameliorates osteoarthritis progression through activation of Nrf2/HO-1 and inhibition of the NF- κ B pathway. *International Immunopharmacology* 108:108864 DOI: <https://doi.org/10.1016/j.intimp.2022.108864>.
- Mansor NI, Ling KH, Rosli R, Hassan Z, Adenan MI & Nordin N (2023) Centella asiatica (L.) Urban. Attenuates Cell Damage in Hydrogen Peroxide-Induced Oxidative Stress in Transgenic Murine Embryonic Stem Cell Line-Derived Neural-Like Cells: A Preliminary Study for Potential Treatment of Alzheimer's Disease. *J Alzheimers Dis* 94:S21-s44 DOI: 10.3233/jad-221233.
- Maramaldi G, Togni S, Franceschi F & Lati E (2013) Anti-inflammaging and antiglycation activity of a novel botanical ingredient from African biodiversity (Centevita™). *Clin Cosmet Investig Dermatol* 7:1-9 DOI: 10.2147/ccid.S49924.
- Marín-Aguilar F, Lechuga-Vieco AV, Alcocer-Gómez E, Castejón-Vega B, Lucas J, Garrido C, Peralta-García A, Pérez-Pulido AJ, Varela-López A, Quiles JL, Ryffel B, Flores I, Bullón P, Ruiz-Cabello J & Cordero MD (2020) NLRP3 inflammasome suppression improves longevity and prevents cardiac aging in male mice. *Aging Cell* 19:e13050 DOI: 10.1111/acer.13050.
- Maruthiyodan S, Munegowda G, Thomas M, Gangadharan G, Joshi MB, Mumbreakar KD, Goyal A, Mahadevan A, Erayur Mana A, Thrikovil Sankaran M, Marthanda Varma Sankaran V & Kanive Parashiva G (2026) Multiomics analysis reveals early administration of the Medhya rasayana formulation of Bacopa monnieri (L.) Wettst. and Centella asiatica (L.) Urb. prevents neuroinflammation and cognitive deficits in an A β O-injected AD mouse model. *Journal of Ethnopharmacology* 355:120633 DOI: <https://doi.org/10.1016/j.jep.2025.120633>.
- Matthews DG, Khorani M, Bobe G, Caruso M, Magana AA, Gray NE, Quinn JF, Stevens JF, Maier CS & Soumyanath A (2024) Centella asiatica improves cognitive function and alters the hippocampal metabolome of aged Tg2576 and wild-type mice. *J Alzheimers Dis Rep* 8:1611-38 DOI: 10.1177/25424823241296740.
- McGEER PL & McGEER EG (2004) Inflammation and the Degenerative Diseases of Aging. *Annals of the New York Academy of Sciences* 1035:104-16 DOI: <https://doi.org/10.1196/annals.1332.007>.
- Md Pizar M, Chee BJ, Long I & Osman A (2023) Protective effects of Centella asiatica extract on spatial memory and learning deficits in animal model of systemic inflammation induced by lipopolysaccharide. *Ann Med* 55:2224970 DOI: 10.1080/07853890.2023.2224970.

- Miller KN, Victorelli SG, Salmonowicz H, Dasgupta N, Liu T, Passos JF & Adams PD (2021) Cytoplasmic DNA: sources, sensing, and role in aging and disease. *Cell* 184:5506-26 DOI: 10.1016/j.cell.2021.09.034.
- Mittelbrunn M & Kroemer G (2021) Hallmarks of T cell aging. *Nature Immunology* 22:687-98 DOI: 10.1038/s41590-021-00927-z.
- Moldovan M, Lahmar A, Bogdan C, Părăuan S, Tomuță I & Crișan M (2017) Formulation and evaluation of a water-in-oil cream containing herbal active ingredients and ferulic acid. *Chujul Med* 90:212-9 DOI: 10.15386/cjmed-668.
- Monestime O, Davis BA, Layman C, Wheeler KJ, Hack W, Zweig JA, Soumyanath A, Carbone L & Gray NE (2025) Peripheral Blood DNA Methylation Changes in Response to Centella asiatica Treatment in Aged Mice. *Biology (Basel)* 14:DOI: 10.3390/biology14010052.
- Nataraj J, Manivasagam T, Justin Thenmozhi A & Essa MM (2017) Neuroprotective effect of asiatic acid on rotenone-induced mitochondrial dysfunction and oxidative stress-mediated apoptosis in differentiated SH-SY5Y cells. *Nutritional Neuroscience* 20:351-9 DOI: 10.1080/1028415X.2015.1135559.
- Nataraj J, Manivasagam T, Justin Thenmozhi A & Essa MM (2017) Neurotrophic Effect of Asiatic acid, a Triterpene of Centella asiatica Against Chronic 1-Methyl 4-Phenyl 1, 2, 3, 6-Tetrahydropyridine Hydrochloride/Probenecid Mouse Model of Parkinson's disease: The Role of MAPK, PI3K-Akt-GSK3 β and mTOR Signalling Pathways. *Neurochemical Research* 42:1354-65 DOI: 10.1007/s11064-017-2183-2.
- Ni Y, Pan Y, Zhou J, Du H, Yang W, Zheng L, Zheng Y, Jin H, Fu Z, Cai C, He Q & Jin J (2025) Asiaticoside and asiatic acid improve diabetic nephropathy by restoring podocyte autophagy and improving gut microbiota dysbiosis. *Biochemical Pharmacology* 241:117161 DOI: <https://doi.org/10.1016/j.bcp.2025.117161>.
- Niccoli T & Partridge L (2012) Ageing as a Risk Factor for Disease. *Current Biology* 22:R741-R52 DOI: 10.1016/j.cub.2012.07.024.
- Nurhapsari A, Cilmiaty R, Prayitno A, Purwanto B & Soetrisno S (2023) The Role of Asiatic Acid in Preventing Dental Pulp Inflammation: An in-vivo Study. *Clin Cosmet Investig Dent* 15:109-19 DOI: 10.2147/ccide.S408158.
- Omar N, Lokanathan Y, Mohd Razi ZR & Bt Haji Idrus R (2019) The effects of Centella asiatica (L.) Urban on neural differentiation of human mesenchymal stem cells in vitro. *BMC Complement Altern Med* 19:167 DOI: 10.1186/s12906-019-2581-x.
- Papaiahgari S, Zhang Q, Kleeberger SR, Cho H-Y & Reddy SP (2006) Hyperoxia Stimulates an Nrf2-ARE Transcriptional Response via ROS-EGFR-PI3K-Akt/ERK MAP Kinase Signaling in Pulmonary Epithelial Cells. *Antioxidants & Redox Signaling* 8:43-52 DOI: 10.1089/ars.2006.8.43.
- Park S, Park HW, Seo DB, Yoo DS & Bae S (2024) In vitro hair growth-promoting effects of araliadiol via the p38/PPAR- γ signaling pathway in human hair follicle stem cells and dermal papilla cells. *Front Pharmacol* 15:1482898 DOI: 10.3389/fphar.2024.1482898.
- Petr MA, Tulika T, Carmona-Marin LM & Scheibye-Knudsen M (2020) Protecting the Aging Genome. *Trends in Cell Biology* 30:117-32 DOI: 10.1016/j.tcb.2019.12.001.
- Ponnusamy K, Mohan M & Nagaraja HS (2008) Protective antioxidant effect of Centella asiatica bioflavonoids on lead acetate induced neurotoxicity. *Med J Malaysia* 63 Suppl A:102.
- Phoemsapthawee J, Anmawat W, Prasertsri P, Sathalalai P & Leelayuwat N (2022) Does Gotu kola supplementation improve cognitive function, inflammation, and oxidative stress more than multicomponent exercise alone? - a randomized controlled study. *J Exerc Rehabil* 18:330-42 DOI: 10.12965/jer.2244388.194.
- Qi J-J, Li X-X, Zhang Y, Diao Y-F, Hu W-Y, Wang D-L, Jiang H, Zhang J-B, Sun B-X & Liang S (2021) Supplementation with asiatic acid during in vitro maturation improves

- porcine oocyte developmental competence by regulating oxidative stress. *Theriogenology* 172:169-77 DOI: <https://doi.org/10.1016/j.theriogenology.2021.06.013>.
- Qi Z, Ci X, Huang J, Liu Q, Yu Q, Zhou J & Deng X (2017) Asiatic acid enhances Nrf2 signaling to protect HepG2 cells from oxidative damage through Akt and ERK activation. *Biomedicine & Pharmacotherapy* 88:252-9 DOI: <https://doi.org/10.1016/j.biopha.2017.01.067>.
- Qian Y, Xin Z, Lv Y, Wang Z, Zuo L, Huang X, Li Y & Xin H-B (2018) Asiatic acid suppresses neuroinflammation in BV2 microglia via modulation of the Sirt1/NF- κ B signaling pathway. *Food & Function* 9:1048-57 DOI: 10.1039/C7FO01442B.
- Qiu F, Yuan Y, Luo W, Gong YS, Zhang ZM, Liu ZM & Gao L (2022) Asiatic acid alleviates ischemic myocardial injury in mice by modulating mitophagy- and glycolysis-based energy metabolism. *Acta Pharmacol Sin* 43:1395-407 DOI: 10.1038/s41401-021-00763-9.
- Qiu J, Yu L, Zhang X, Wu Q, Wang D, Wang X, Xia C & Feng H (2015) Asiaticoside attenuates lipopolysaccharide-induced acute lung injury via down-regulation of NF- κ B signaling pathway. *International Immunopharmacology* 26:181-7 DOI: <https://doi.org/10.1016/j.intimp.2015.03.022>.
- Quiles J, Cabrera M, Jones J, Tsapekos M & Caturla N (2022) In Vitro Determination of the Skin Anti-Aging Potential of Four-Component Plant-Based Ingredient. *Molecules* 27:DOI: 10.3390/molecules27228101.
- Ramesh BN, Girish TK, Raghavendra RH, Naidu KA, Rao UJ & Rao KS (2014) Comparative study on anti-oxidant and anti-inflammatory activities of Caesalpinia crista and Centella asiatica leaf extracts. *J Pharm Bioallied Sci* 6:86-91 DOI: 10.4103/0975-7406.129172.
- Ratray NJW, Trivedi DK, Xu Y, Chandola T, Johnson CH, Marshall AD, Mekli K, Ratray Z, Tampubolon G, Vanhoutte B, White IR, Wu FCW, Pendleton N, Nazroo J & Goodacre R (2019) Metabolic dysregulation in vitamin E and carnitine shuttle energy mechanisms associate with human frailty. *Nat Commun* 10:5027 DOI: 10.1038/s41467-019-12716-2.
- Rather MA, Thenmozhi AJ, Manivasagam T, Bharathi MD, Essa MM & Guillemain GJ (2018) Neuroprotective role of Asiatic acid in aluminium chloride induced rat model of Alzheimer's disease. *Front Biosci (Schol Ed)* 10:262-75 DOI: 10.2741/s514.
- Reid Thompson W, Hornby B, Manuel R, Bradley E, Laux J, Carr J & Vernon HJ (2021) A phase 2/3 randomized clinical trial followed by an open-label extension to evaluate the effectiveness of elamipretide in Barth syndrome, a genetic disorder of mitochondrial cardiolipin metabolism. *Genet Med* 23:471-8 DOI: 10.1038/s41436-020-01006-8.
- Ren L, Cao Q-X, Zhai F-R, Yang S-Q & Zhang H-X (2016) Asiatic acid exerts anticancer potential in human ovarian cancer cells via suppression of PI3K/Akt/mTOR signalling. *Pharmaceutical Biology* 54:2377-82 DOI: 10.3109/13880209.2016.1156709.
- Reynolds JC, Lai RW, Woodhead JST, Joly JH, Mitchell CJ, Cameron-Smith D, Lu R, Cohen P, Graham NA, Benayoun BA, Merry TL & Lee C (2021) MOTS-c is an exercise-induced mitochondrial-encoded regulator of age-dependent physical decline and muscle homeostasis. *Nat Commun* 12:470 DOI: 10.1038/s41467-020-20790-0.
- Ridker PM, MacFadyen JG, Thuren T, Everett BM, Libby P, Glynn RJ, Ridker P, Lorenzatti A, Krum H, Varigos J, Siostrzonek P, Sinnaeve P, Fonseca F, Nicolau J, Gotcheva N, Genest J, Yong H, Urina-Triana M, Milicic D, Cifkova R, Vettus R, Koenig W, Anker SD, Manolis AJ, Wyss F, Forster T, Sigurdsson A, Pais P, Fucili A, Ogawa H, Shimokawa H, Veze I, Petrauskiene B, Salvador L, Kastelein J, Cornel JH, Klemsdal TO, Medina F, Budaj A, Vida-Simiti L, Kobalava Z, Otasevic P, Pella D, Lainscak M, Seung K-B, Commerford P, Dellborg M, Donath M, Hwang J-J, Kultursay H, Flather M, Ballantyne C, Bilazarian S, Chang W, East C, Everett B, Forgosh L, Glynn R, Harris B, Libby P, Ligueros M, Thuren T, Bohula E, Charmarathi B, Cheng S, Chou S, Danik J, McMahon G, Maron B, Ning M, Olenchock B, Pande R, Perlstein T, Pradhan A, Rost N, Singhal

- A, Taqueti V, Wei N, Burris H, Cioffi A, Dalseg AM, Ghosh N, Gralow J, Mayer T, Rugo H, Fowler V, Limaye AP, Cosgrove S, Levine D, Lopes R, Scott J, Thuren T, Ligueros M, Hilkert R, Tamesby G, Mickel C, Manning B, Woelcke J, Tan M, Manfredda S, Ponce T, Kam J, Saini R, Banker K, Salko T, Nandy P, Tawfik R, O'Neil G, Manne S, Jirvankar P, Lal S, Nema D, Jose J, Collins R, Bailey K, Blumenthal R, Colhoun H, Gersh B & Glynn RJ (2017) Effect of interleukin-1 β inhibition with canakinumab on incident lung cancer in patients with atherosclerosis: exploratory results from a randomised, double-blind, placebo-controlled trial. *The Lancet* 390:1833-42 DOI: 10.1016/S0140-6736(17)32247-X.
- Rowe K, Gray NE, Zweig JA, Law A, Techen N, Maier CS, Soumyanath A & Kretzschmar D (2024) Centella asiatica and its caffeoylquinic acid and triterpene constituents increase dendritic arborization of mouse primary hippocampal neurons and improve age-related locomotion deficits in Drosophila. *Front Aging* 5:1374905 DOI: 10.3389/fragi.2024.1374905.
- Sari DCR, Budiharjo S, Afifah H, Jasmin D, Kokasih O, Putri TG, Arifiani K, Setyaningsih WAW & Arfian N (2021) Centella asiatica Extract Attenuates Kidney Fibrosis Through Reducing Mesenchymal Transition and Inflammation in Ureteral Ligation Model in Mice. *Front Pharmacol* 12:621894 DOI: 10.3389/fphar.2021.621894.
- Sbrini G, Brivio P, Sangiovanni E, Fumagalli M, Racagni G, Dell'Agli M & Calabrese F (2020) Chronic Treatment with a Phytosomal Preparation Containing Centella asiatica L. and Curcuma longa L. Affects Local Protein Synthesis by Modulating the BDNF-mTOR-S6 Pathway. *Biomedicines* 8:DOI: 10.3390/biomedicines8120544.
- Sciorati C, Gamberale R, Monno A, Citterio L, Lanzani C, De Lorenzo R, Ramirez GA, Esposito A, Manunta P, Manfredi AA & Rovere-Querini P (2020) Pharmacological blockade of TNF α prevents sarcopenia and prolongs survival in aging mice. *Aging (Albany NY)* 12:23497-508 DOI: 10.18632/aging.202200.
- Schmauck-Medina T, Molière A, Lautrup S, Zhang J, Chlopicki S, Madsen HB, Cao S, Soendenbroe C, Mansell E, Vestergaard MB, Li Z, Shiloh Y, Opresko PL, Egly JM, Kirkwood T, Verdin E, Bohr VA, Cox LS, Stevnsner T, Rasmussen LJ & Fang EF (2022) New hallmarks of ageing: a 2022 Copenhagen ageing meeting summary. *Aging (Albany NY)* 14:6829-39 DOI: 10.18632/aging.204248.
- Shen H, Zhu F, Li J, Tang S, Zhang Y & Zhang J (2020) Protective Effect of Asiaticoside on Radiation-induced Proliferation Inhibition and DNA Damage of Fibroblasts and Mice Death. *Open Life Sci* 15:145-51 DOI: 10.1515/biol-2020-0015.
- Shen X, Guo M, Yu H, Liu D, Lu Z & Lu Y (2019) Propionibacterium acnes related anti-inflammation and skin hydration activities of madecassoside, a pentacyclic triterpene saponin from Centella asiatica. *Bioscience, Biotechnology, and Biochemistry* 83:561-8 DOI: 10.1080/09168451.2018.1547627.
- Shin HY, Jeong YY, Kim J-E, Shin K-S & Yu K-W (2026) Anti-colitic effects of Centella asiatica (L.) Urb. juice via ERK/p38 and NF- κ B signaling modulation and the characterization of a key marker compound. *Journal of Ethnopharmacology* 355:120657 DOI: <https://doi.org/10.1016/j.jep.2025.120657>.
- Shinomol GK & Muralidhara (2008) Effect of Centella asiatica leaf powder on oxidative markers in brain regions of prepubertal mice in vivo and its in vitro efficacy to ameliorate 3-NPA-induced oxidative stress in mitochondria. *Phytomedicine* 15:971-84 DOI: <https://doi.org/10.1016/j.phymed.2008.04.010>.
- Shlisky J, Bloom DE, Beaudreault AR, Tucker KL, Keller HH, Freund-Levi Y, Fielding RA, Cheng FW, Jensen GL, Wu D & Meydani SN (2017) Nutritional Considerations for Healthy Aging and Reduction in Age-Related Chronic Disease. *Advances in Nutrition* 8:17-26 DOI: <https://doi.org/10.3945/an.116.013474>.
- Smith LK, He Y, Park JS, Bieri G, Snelthage CE, Lin K, Gontier G, Wabl R, Plambeck KE, Udeochu J, Wheatley EG, Bouchard J, Eggel A, Narasimha R, Grant JL, Luo J, Wyss-

- Coray T & Villeda SA (2015) β 2-microglobulin is a systemic pro-aging factor that impairs cognitive function and neurogenesis. *Nat Med* 21:932-7 DOI: 10.1038/nm.3898.
- Somasundaram I, Jain SM, Blot-Chabaud M, Pathak S, Banerjee A, Rawat S, Sharma NR & Duttaroy AK (2024) Mitochondrial dysfunction and its association with age-related disorders. *Front Physiol* 15:1384966 DOI: 10.3389/fphys.2024.1384966.
- Songvut P, Chariyavilaskul P, Khemawoot P & Tansawat R (2021) Pharmacokinetics and metabolomics investigation of an orally modified formula of standardized *Centella asiatica* extract in healthy volunteers. *Sci Rep* 11:6850 DOI: 10.1038/s41598-021-86267-2.
- Speers AB, Wright KM, Brandes MS, Kedjejian N, Matthews DG, Caruso M, Harris CJ, Koike S, Nguyen T, Quinn JF, Soumyanath A & Gray NE (2024) Mode of administration influences plasma levels of active *Centella asiatica* compounds in 5xFAD mice while markers of neuroinflammation remain unaltered. *Front Neurosci* 18:1277626 DOI: 10.3389/fnins.2024.1277626.
- Subathra M, Shila S, Devi MA & Panneerselvam C (2005) Emerging role of *Centella asiatica* in improving age-related neurological antioxidant status. *Experimental Gerontology* 40:707-15 DOI: <https://doi.org/10.1016/j.exger.2005.06.001>.
- Sun B, Wu L, Wu Y, Zhang C, Qin L, Hayashi M, Kudo M, Gao M & Liu T (2020) Therapeutic Potential of *Centella asiatica* and Its Triterpenes: A Review. *Front Pharmacol* 11:568032 DOI: 10.3389/fphar.2020.568032.
- Sung Y-Y, Son E, Kim D-S, Kim S-H, Yang W-K & Kim M (2025) Lactobacillus-Fermented *Centella asiatica* Extract Inhibits Airway Inflammation in Cigarette Smoke Extract/LPS-Induced Mice. *Plants* 14:3416.
- Tan SC, Bhattamisra SK, Chellappan DK & Candasamy M (2021) Actions and Therapeutic Potential of Madecassoside and Other Major Constituents of *Centella asiatica*: A Review. *Applied Sciences* 11:8475.
- Tang X-L, Yang X-Y, Jung H-J, Kim S-Y, Jung S-Y, Choi D-Y, Park W-C & Park H (2009) Asiatic Acid Induces Colon Cancer Cell Growth Inhibition and Apoptosis through Mitochondrial Death Cascade. *Biological and Pharmaceutical Bulletin* 32:1399-405 DOI: 10.1248/bpb.32.1399.
- Tsoukalas D, Zlatian O, Mitroi M, Renieri E, Tsatsakis A, Izotov BN, Burada F, Sosoi S, Burada E, Buga AM, Rogoveanu I, Docea AO & Calina D (2021) A Novel Nutraceutical Formulation Can Improve Motor Activity and Decrease the Stress Level in a Murine Model of Middle-Age Animals. *J Clin Med* 10:DOI: 10.3390/jcm10040624.
- Thannium S, Laomeephol C, Pavasant P, Osathanon T, Tabata Y, Wang C & Luckanagul JA (2023) Osteogenic induction of asiatic acid derivatives in human periodontal ligament stem cells. *Sci Rep* 13:14102 DOI: 10.1038/s41598-023-41388-8.
- Umka Welbat J, Sirichoat A, Chaijaroonkhanarak W, Prachaney P, Pannangrong W, Pakdeechote P, Sripanidkulchai B & Wigmore P (2016) Asiatic Acid Prevents the Deleterious Effects of Valproic Acid on Cognition and Hippocampal Cell Proliferation and Survival. *Nutrients* 8:DOI: 10.3390/nu8050303.
- United Nations DoEaSA, Population Division (2013) World Population Ageing 2013.
- Valletta S, Thomas A, Meng Y, Ren X, Drissen R, Sengül H, Di Genua C & Nerlov C (2020) Micro-environmental sensing by bone marrow stroma identifies IL-6 and TGF β 1 as regulators of hematopoietic ageing. *Nat Commun* 11:4075 DOI: 10.1038/s41467-020-17942-7.
- Vijg J & Dong X (2020) Pathogenic Mechanisms of Somatic Mutation and Genome Mosaicism in Aging. *Cell* 182:12-23 DOI: 10.1016/j.cell.2020.06.024.
- Vijg J & Suh Y (2013) Genome Instability and Aging. *Annual Review of Physiology* 75:645-68 DOI: <https://doi.org/10.1146/annurev-physiol-030212-183715>.
- Villeda SA, Luo J, Mosher KI, Zou B, Britschgi M, Bieri G, Stan TM, Fainberg N, Ding Z, Eggel A, Lucin KM, Czirr E, Park JS, Couillard-Després S, Aigner L, Li G, Peskind ER, Kaye JA, Quinn JF, Galasko DR, Xie XS, Rando TA & Wyss-Coray T (2011) The ageing

- systemic milieu negatively regulates neurogenesis and cognitive function. *Nature* 477:90-4 DOI: 10.1038/nature10357.
- Wan J, Gong X, Jiang R, Zhang Z & Zhang L (2013) Antipyretic and Anti-inflammatory Effects of Asiaticoside in Lipopolysaccharide-treated Rat through Up-regulation of Heme Oxygenase-1. *Phytotherapy Research* 27:1136-42 DOI: <https://doi.org/10.1002/ptr.4838>.
- Wang ZH (2014) Anti-glycative effects of asiatic acid in human keratinocyte cells. *Biomedicine (Taipei)* 4:19 DOI: 10.7603/s40681-014-0019-9.
- Wei L, Chen Q, Guo A, Fan J, Wang R & Zhang H (2018) Asiatic acid attenuates CCl4-induced liver fibrosis in rats by regulating the PI3K/AKT/mTOR and Bcl-2/Bax signaling pathways. *International Immunopharmacology* 60:1-8 DOI: <https://doi.org/10.1016/j.intimp.2018.04.016>.
- Widgerow A, Grivet-Seyve M, Anjuwon S, Emesiani C & Meckfessel M (2025) Efficacy and Tolerability of a Cream in Aging Skin. *J Drugs Dermatol* 24:524-9 DOI: 10.36849/jdd.9025.
- Won J-H, Shin J-S, Park H-J, Jung H-J, Koh D-J, Jo B-G, Lee J-Y, Yun K & Lee K-T (2010) Anti-inflammatory Effects of Madecassic Acid via the Suppression of NF- κ B Pathway in LPS-Induced RAW 264.7 Macrophage Cells. *Planta Med* 76:251-7 DOI: 10.1055/s-0029-1186142.
- Xiao F, Li Q, Zeng W, Tang B, Chen Q, Wu C, Duan Z, Chen H, Rui S & Liu B (2025) Centella asiatica enhances diabetic wound healing by decreasing macrophage-driven inflammation via the AKT/MAPK/NF- κ B pathway. *Front Pharmacol* 16:1632573 DOI: 10.3389/fphar.2025.1632573.
- Xu MF, Xiong YY, Liu JK, Qian JJ, Zhu L & Gao J (2012) Asiatic acid, a pentacyclic triterpene in Centella asiatica, attenuates glutamate-induced cognitive deficits in mice and apoptosis in SH-SY5Y cells. *Acta Pharmacol Sin* 33:578-87 DOI: 10.1038/aps.2012.3.
- Xu P, Wang M, Song WM, Wang Q, Yuan GC, Sudmant PH, Zare H, Tu Z, Orr ME & Zhang B (2022) The landscape of human tissue and cell type specific expression and coregulation of senescence genes. *Mol Neurodegener* 17:5 DOI: 10.1186/s13024-021-00507-7.
- Xu X, Wang Y, Wei Z, Wei W, Zhao P, Tong B, Xia Y & Dai Y (2017) Madecassic acid, the contributor to the anti-colitis effect of madecassoside, enhances the shift of Th17 toward Treg cells via the PPAR γ /AMPK/ACC1 pathway. *Cell Death Dis* 8:e2723 DOI: 10.1038/cddis.2017.150.
- Xu Y & Wan W (2023) Acetylation in the regulation of autophagy. *Autophagy* 19:379-87 DOI: 10.1080/15548627.2022.2062112.
- Yang C, Guo Y, Huang T-s, Zhao J, Huang X-J, Tang H-x, An N, Pan Q, Xu Y-z & Liu H-f (2018) Asiatic acid protects against cisplatin-induced acute kidney injury via anti-apoptosis and anti-inflammation. *Biomedicine & Pharmacotherapy* 107:1354-62 DOI: <https://doi.org/10.1016/j.biopha.2018.08.126>.
- Yatmark P, Anutagerngkum P, Huajiantug S, Tantisira MH, Ngampramuan S, Svasti S & Morales NP (2025) A standardized extract of Centella asiatica (ECa 233) alleviates brain injury and improves brain function in β -thalassemia mice with iron overload. *BMC Complement Med Ther* 25:412 DOI: 10.1186/s12906-025-05145-w.
- Yi C, Si L, Xu J, Yang J, Wang Q & Wang X (2020) Effect and mechanism of asiatic acid on autophagy in myocardial ischemia-reperfusion injury in vivo and in vitro. *Exp Ther Med* 20:54 DOI: 10.3892/etm.2020.9182.
- Yi C, Song M, Sun L, Si L, Yu D, Li B, Lu P, Wang W & Wang X (2022) Asiatic Acid Alleviates Myocardial Ischemia-Reperfusion Injury by Inhibiting the ROS-Mediated Mitochondria-Dependent Apoptosis Pathway. *Oxid Med Cell Longev* 2022:3267450 DOI: 10.1155/2022/3267450.
- Yuan Y, Zhang H, Sun F, Sun S, Zhu Z & Chai Y (2015) Biopharmaceutical and pharmacokinetic characterization of asiatic acid in Centella asiatica as determined by a

- sensitive and robust HPLC-MS method. *J Ethnopharmacol* 163:31-8 DOI: 10.1016/j.jep.2015.01.006.
- Yun K-J, Kim J-Y, Kim J-B, Lee K-W, Jeong S-Y, Park H-J, Jung H-J, Cho Y-W, Yun K & Lee K-T (2008) Inhibition of LPS-induced NO and PGE2 production by asiatic acid via NF- κ B inactivation in RAW 264.7 macrophages: Possible involvement of the IKK and MAPK pathways. *International Immunopharmacology* 8:431-41 DOI: <https://doi.org/10.1016/j.intimp.2007.11.003>.
- Yuyun X, Xi C, Qing Y, Lin X, Ke R & Bingwei S (2018) Asiatic acid attenuates lipopolysaccharide-induced injury by suppressing activation of the Notch signaling pathway. *Oncotarget* 9:15036-46 DOI: 10.18632/oncotarget.24542.
- Zhang H, Alsaleh G, Feltham J, Sun Y, Napolitano G, Riffelmacher T, Charles P, Frau L, Hublitz P, Yu Z, Mohammed S, Ballabio A, Balabanov S, Mellor J & Simon AK (2019) Polyamines Control eIF5A Hypusination, TFE3 Translation, and Autophagy to Reverse B Cell Senescence. *Mol Cell* 76:110-25.e9 DOI: 10.1016/j.molcel.2019.08.005.
- Zhang Y, Meng X & Liu K (2022) The modulation of cAMP/PKA pathway by asiaticoside ameliorates high glucose-induced inflammation and apoptosis of retinal pigment epithelial cells. *Journal of Bioenergetics and Biomembranes* 54:9-16 DOI: 10.1007/s10863-021-09929-w.
- Zhong W, Zhu G, Lu D, Wang L, Zhang B, Su Z & Qin Z (2025) Madecassoside exerts anti-neurodegeneration effects in protein l-isoaspartyl methyltransferase deficient mice via targeting PRDX2-mediated oxidative stress and inflammation. *Biochemical Pharmacology* 239:117034 DOI: <https://doi.org/10.1016/j.bcp.2025.117034>.
- Zhu Z, Cui L, Yang J, Vong CT, Hu Y, Xiao J, Chan G, He Z & Zhong Z (2021) Anticancer effects of asiatic acid against doxorubicin-resistant breast cancer cells via an AMPK-dependent pathway in vitro. *Phytomedicine* 92:153737 DOI: <https://doi.org/10.1016/j.phymed.2021.153737>.
- Zofia N, Martyna ZD, Aleksandra Z & Tomasz B (2020) Comparison of the Antiaging and Protective Properties of Plants from the Apiaceae Family. *Oxid Med Cell Longev* 2020:5307614 DOI: 10.1155/2020/5307614.
- Zulkipli NN, Zakaria R, Long I, Abdullah SF, Muhammad EF, Wahab HA & Sasongko TH (2020) In Silico Analyses and Cytotoxicity Study of Asiaticoside and Asiatic Acid from Malaysian Plant as Potential mTOR Inhibitors. *Molecules* 25:DOI: 10.3390/molecules25173991.